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# Systems Biology and Evolutionary Computation

GECCO Tutorial

July 8, 2006

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ETH Zurich, Switzerland

**Reverse**  
**Engineering**

**ETH**

Eidgenössische Technische Hochschule Zürich  
Swiss Federal Institute of Technology Zurich

# Goals and Schedule

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## Questions

- What is Systems Biology?
- What are the basic types of biological experiments and measurements?
- What are the computational issues in Systems Biology?
- What are examples of successful application of EC in Systems Biology?

## Schedule

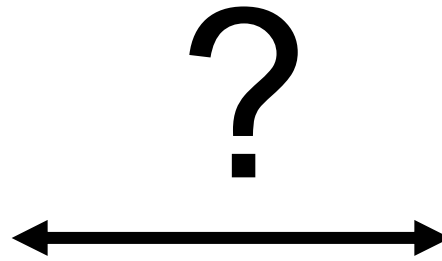
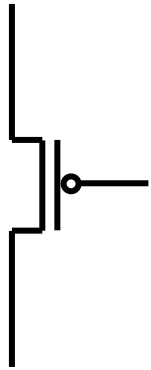
- 50 min Part 1: Introduction to Systems Biology.
- 10 min break
- 50 min Part 2: Computational Issues in Systems Biology.

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# 1. Introduction to Systems Biology

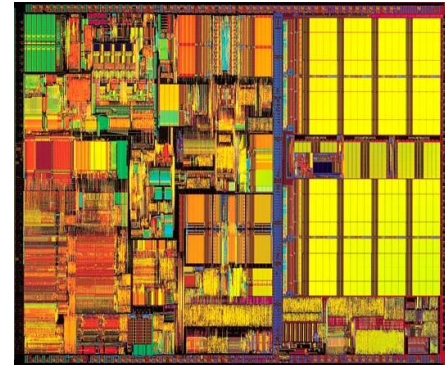
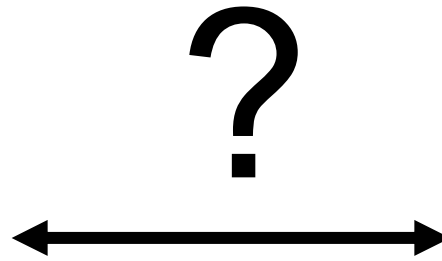
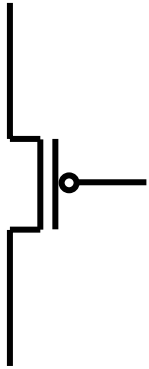
# Reverse Engineering Problem

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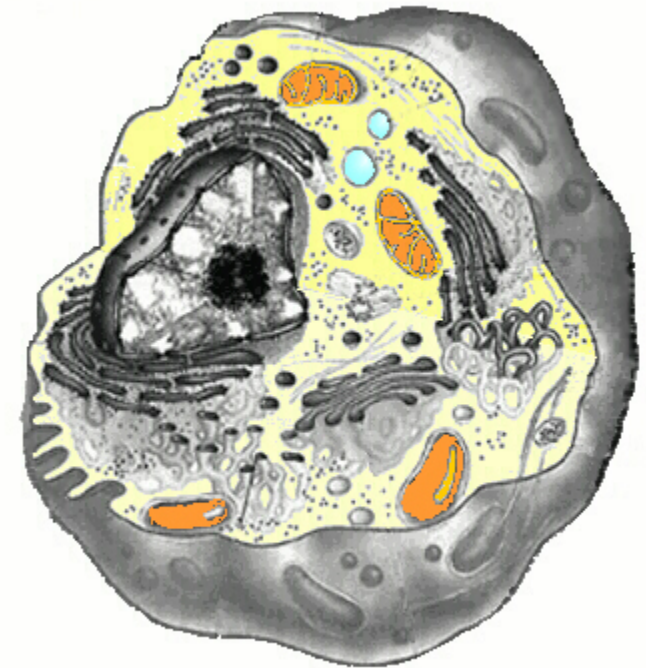
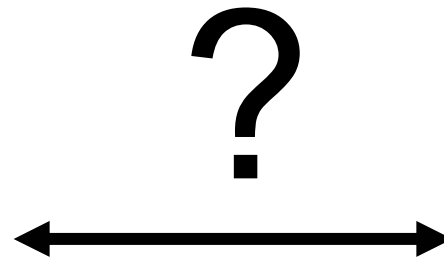
# Reverse Engineering Problem

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# Reverse Engineering Problem II

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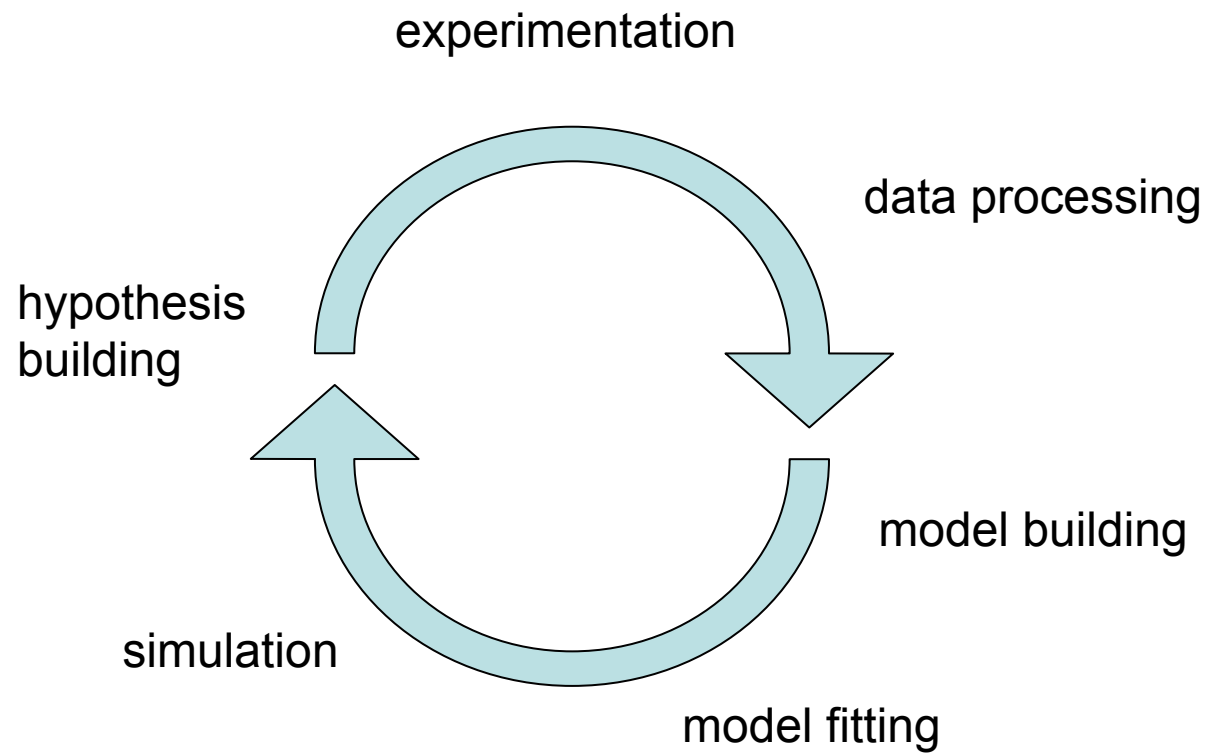
# Systems Biology

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- **A Definition:** Understanding of network behavior using computational approaches tightly linked to experiments. (M. Cassman)
- **Goals:**
  - system level understanding
  - simulators for cells and organisms
  - personalized, predictive and preventive medicine
- **Methods:**
  - experiment: mostly high throughput
  - models
  - computational analysis
- **Key Idea:** capture emergent properties

# Closed Loop Biology

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# Compared to Classical Biology

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## Classical Biology

- focus on single elements (gene, protein, pathway)
- focus on building blocks
- bottom up
- interpretation
  
- Examples:
  - structure determination of a protein
  - study effects of knockout on one pathway

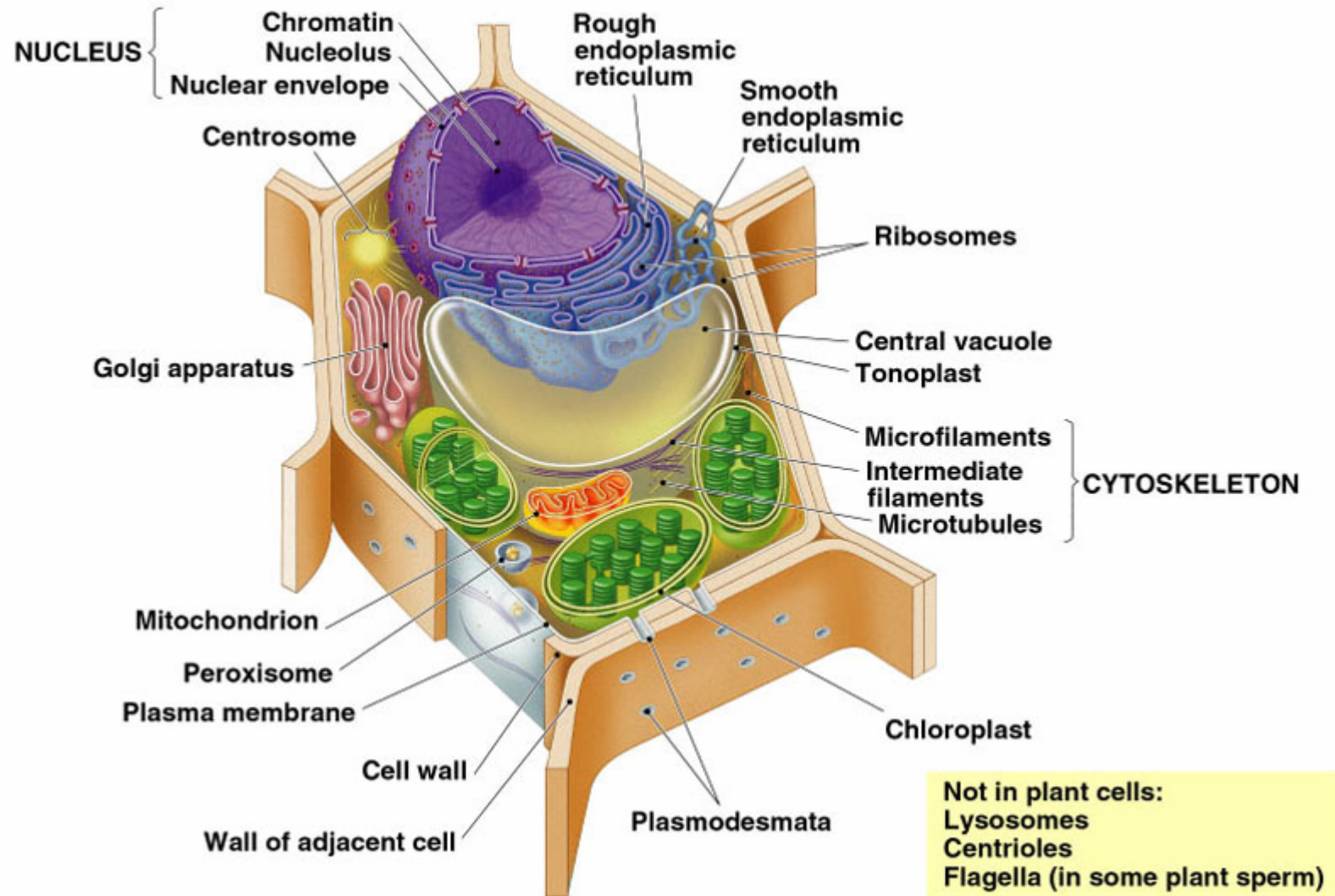
## Systems Biology

- focus on all elements (genome, proteome, metabolome)
- focus on interactions
- top down
- simulation
  
- Examples:
  - module identification
  - robustness analysis of genetic networks

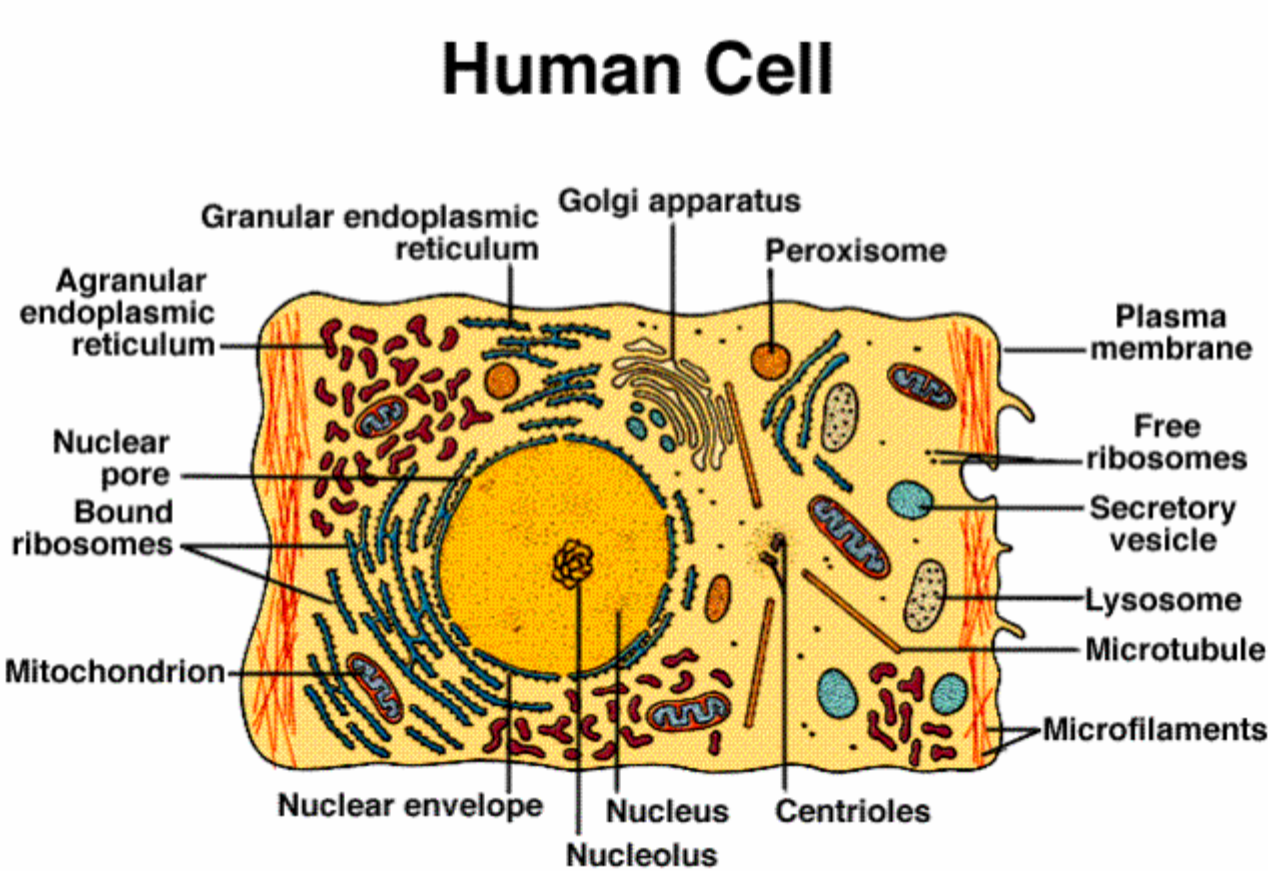
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Some cell biology...

# Cellular Compartments of a Plant Cell



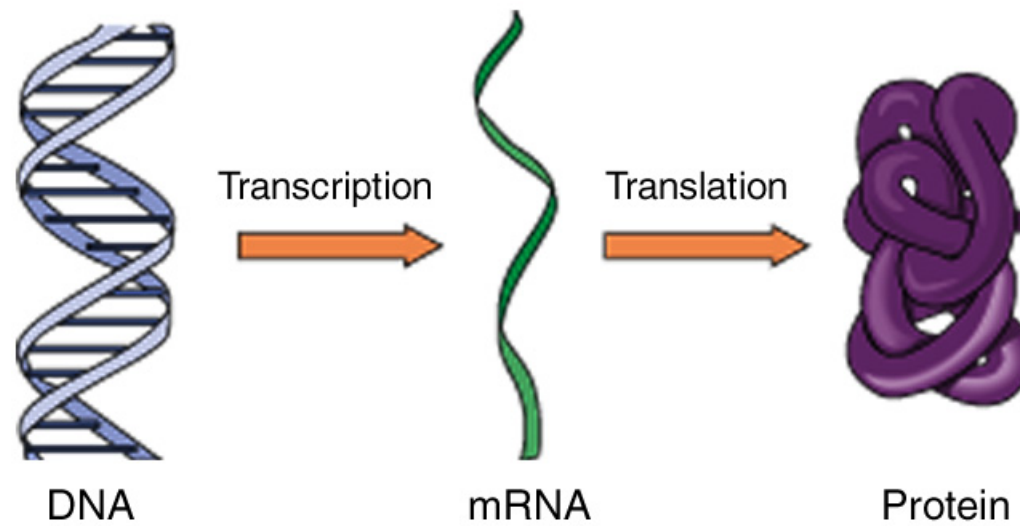
# Cellular Compartments of a Human Cell



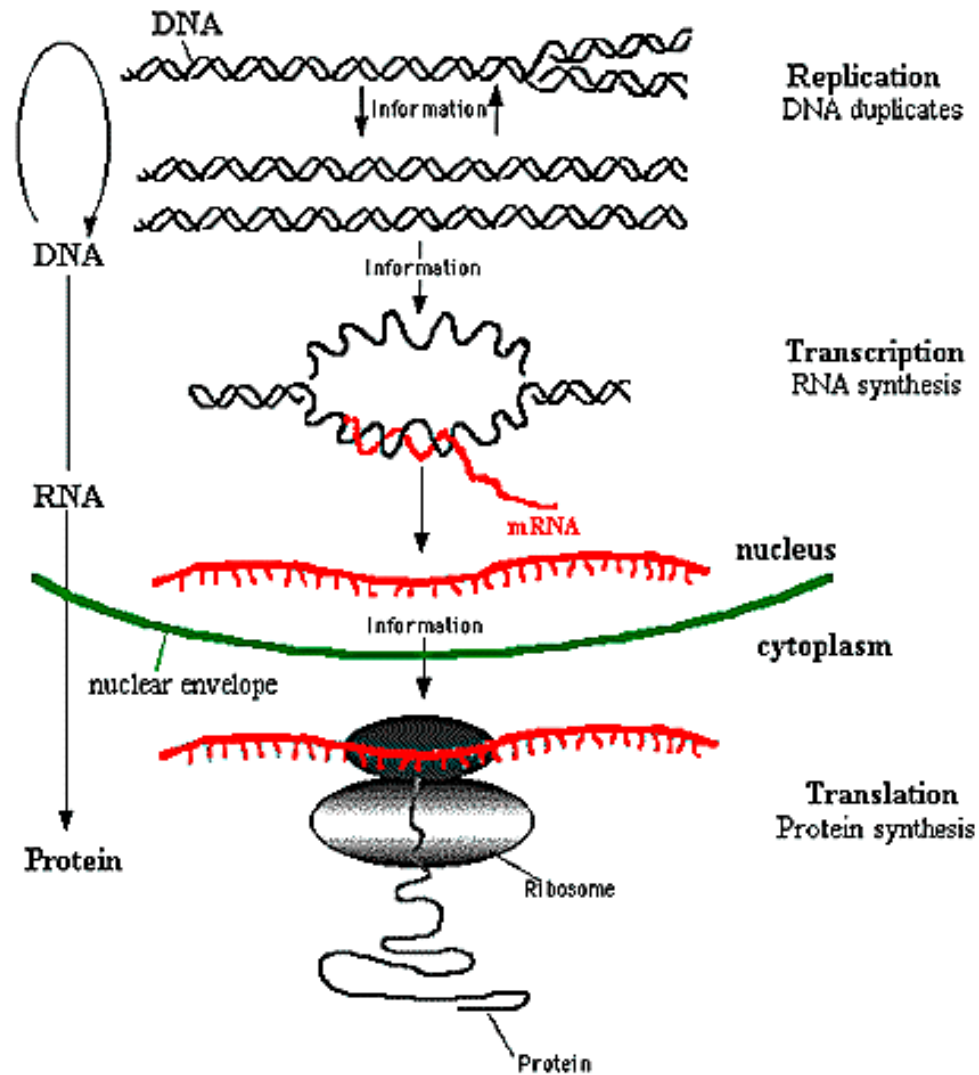
# Central Dogma of Molecular Biology

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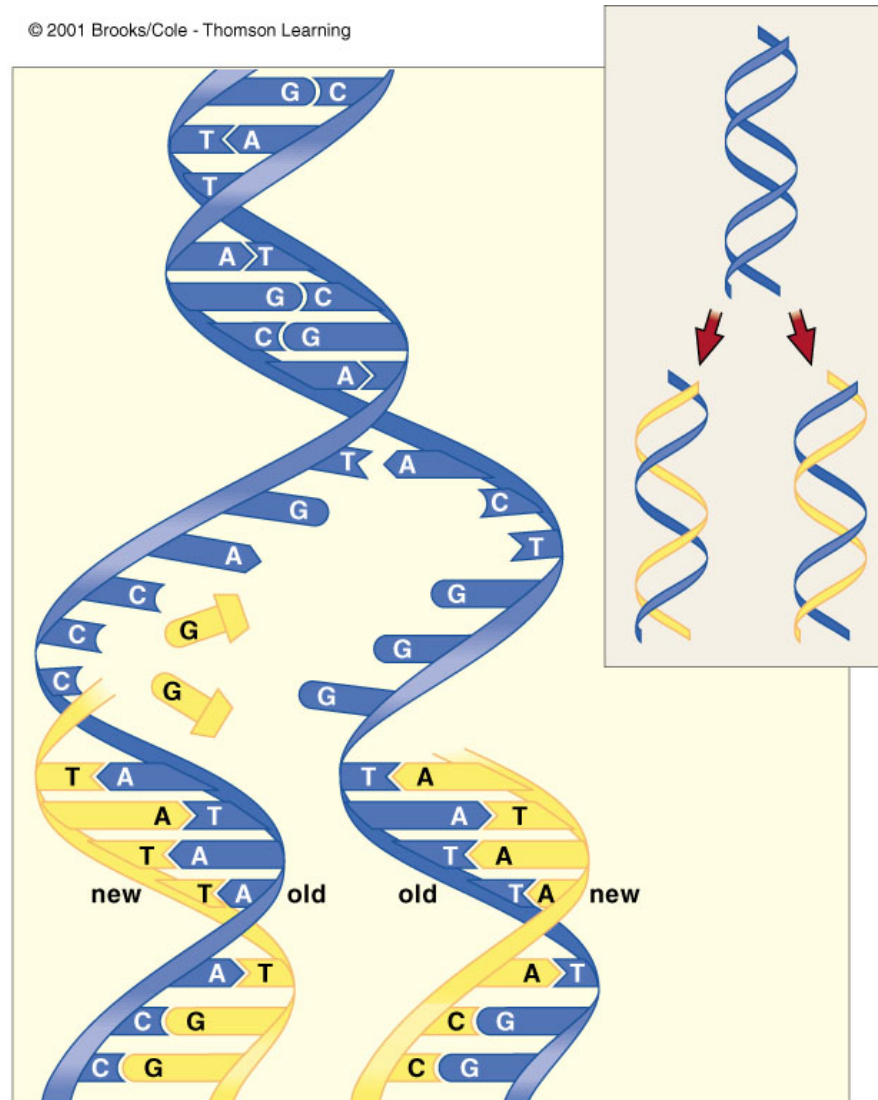
# Central Dogma of Molecular Biology



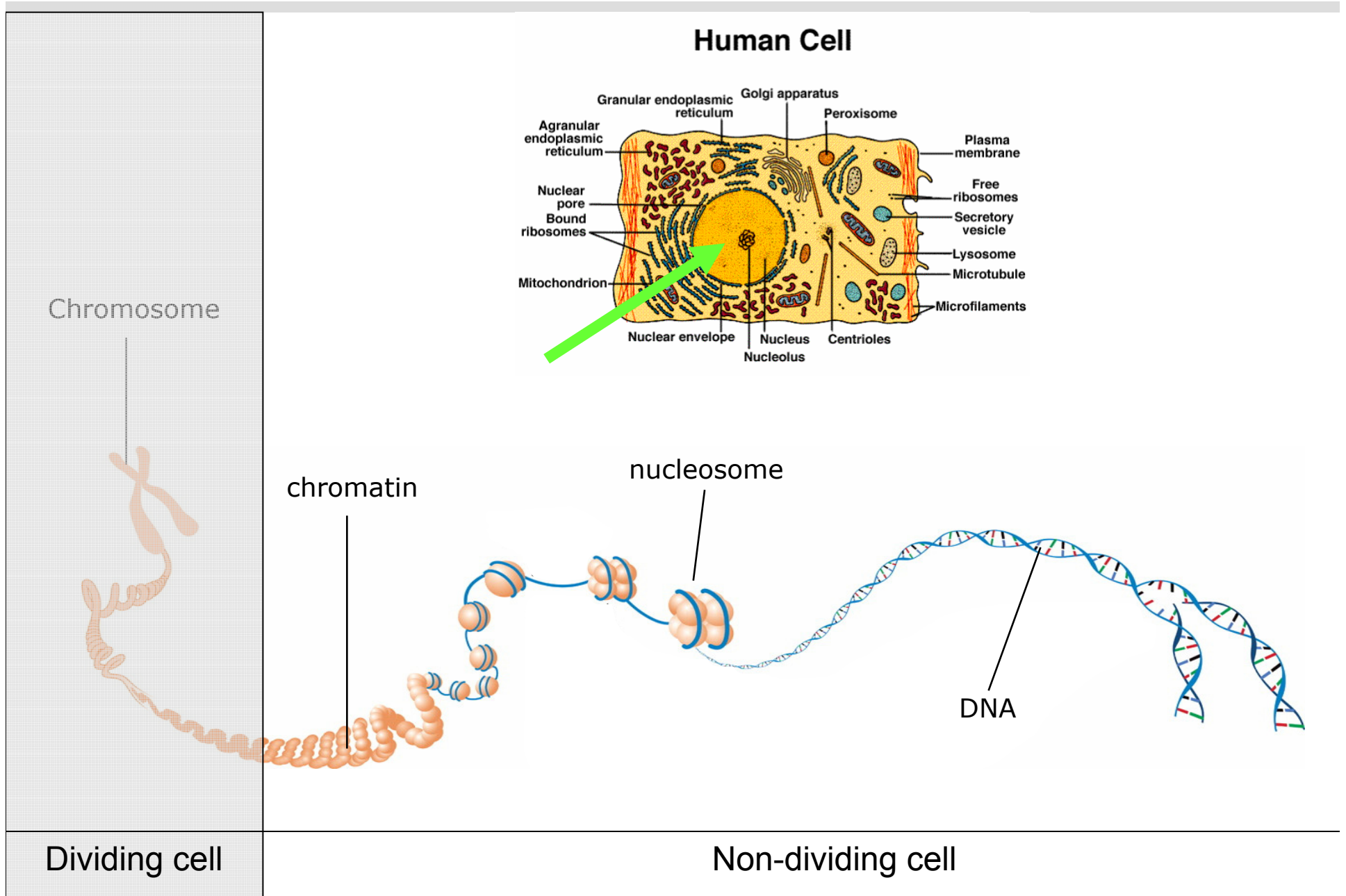
**The Central Dogma of Molecular Biology**

# DNA Replication

© 2011 Brooks/Cole - Thomson Learning

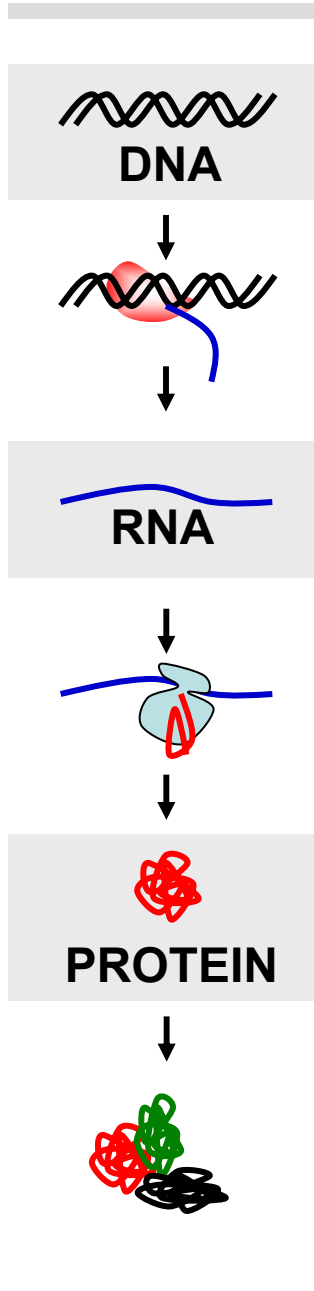


# Chromosome – chromatin – nucleosome – gene

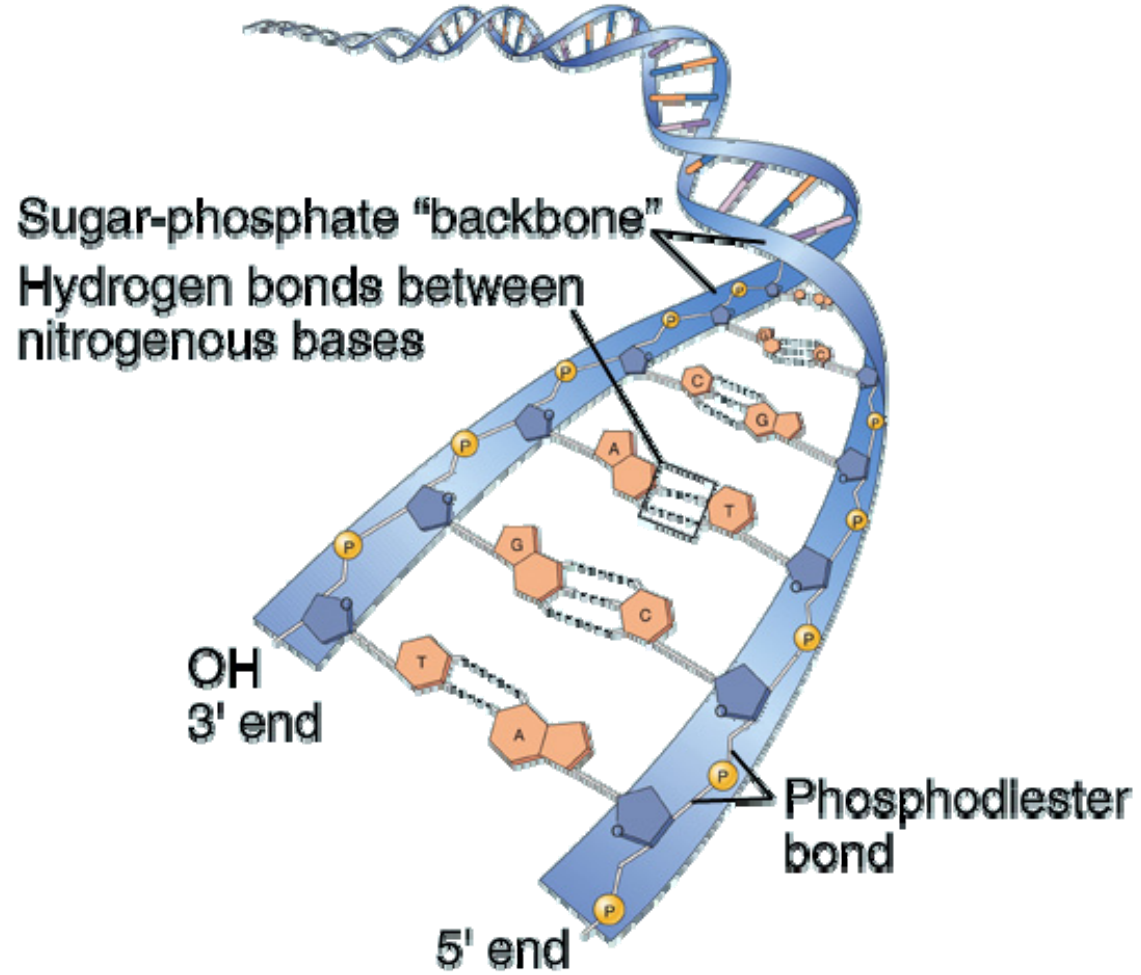




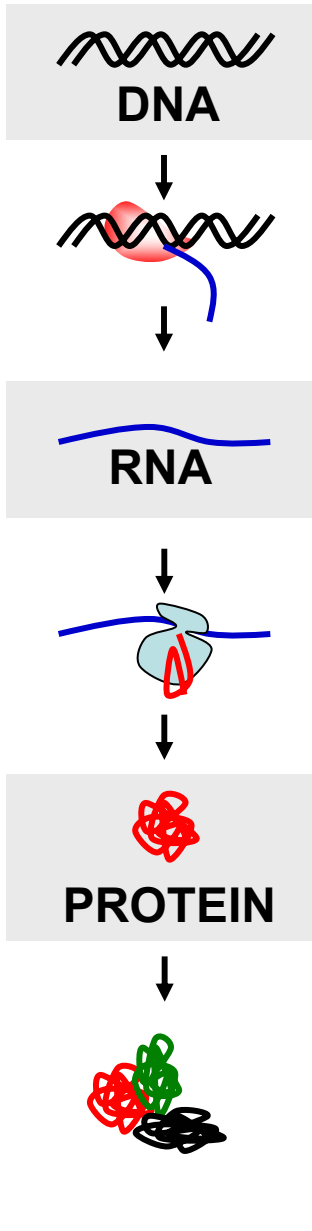
# Genes



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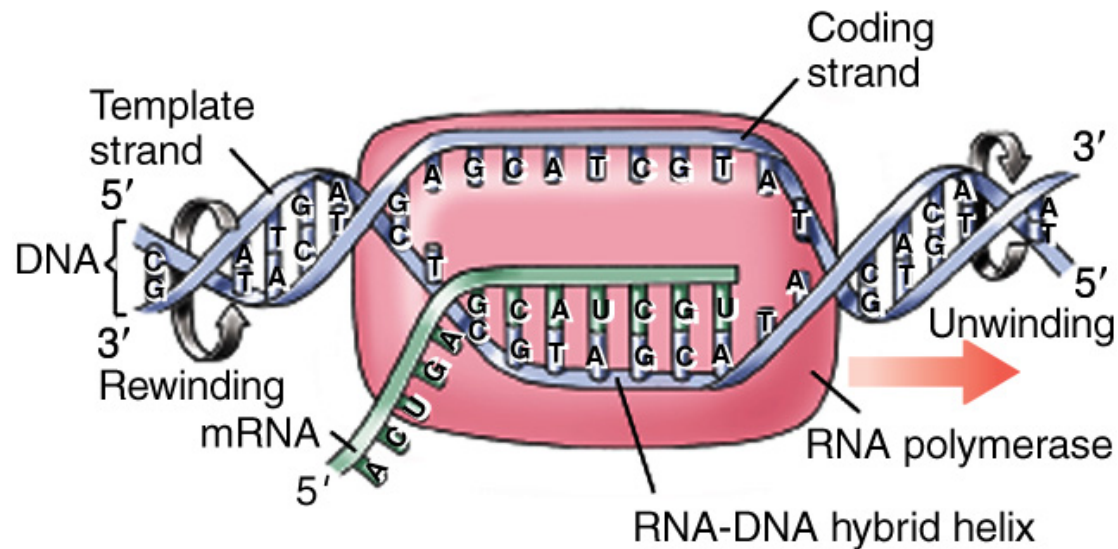


# Transcription

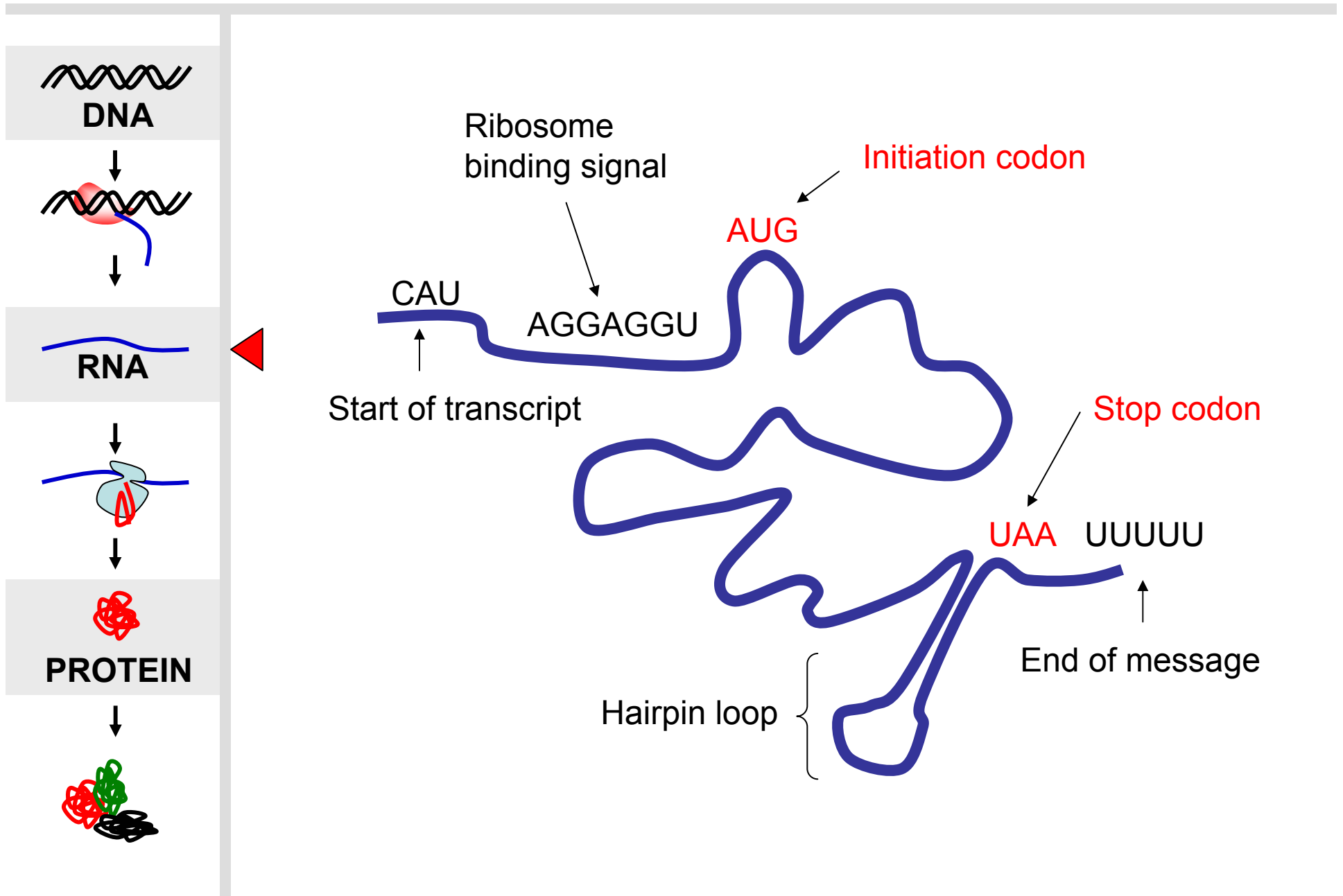


- The process by which a molecule of DNA is copied into a complementary strand of RNA.
- 1 Strand DNA → 2 Strands RNA
- RNA Polymerase

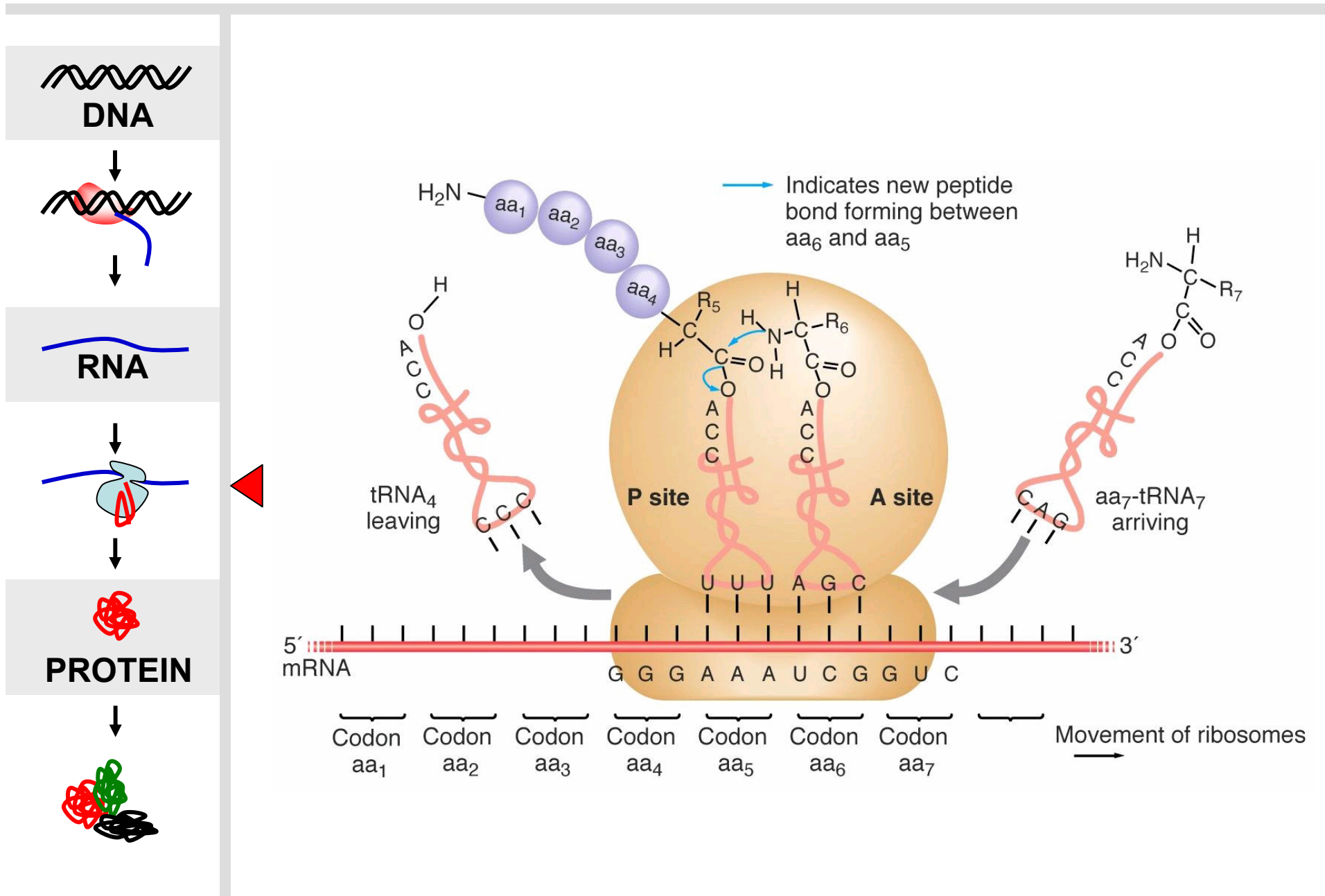
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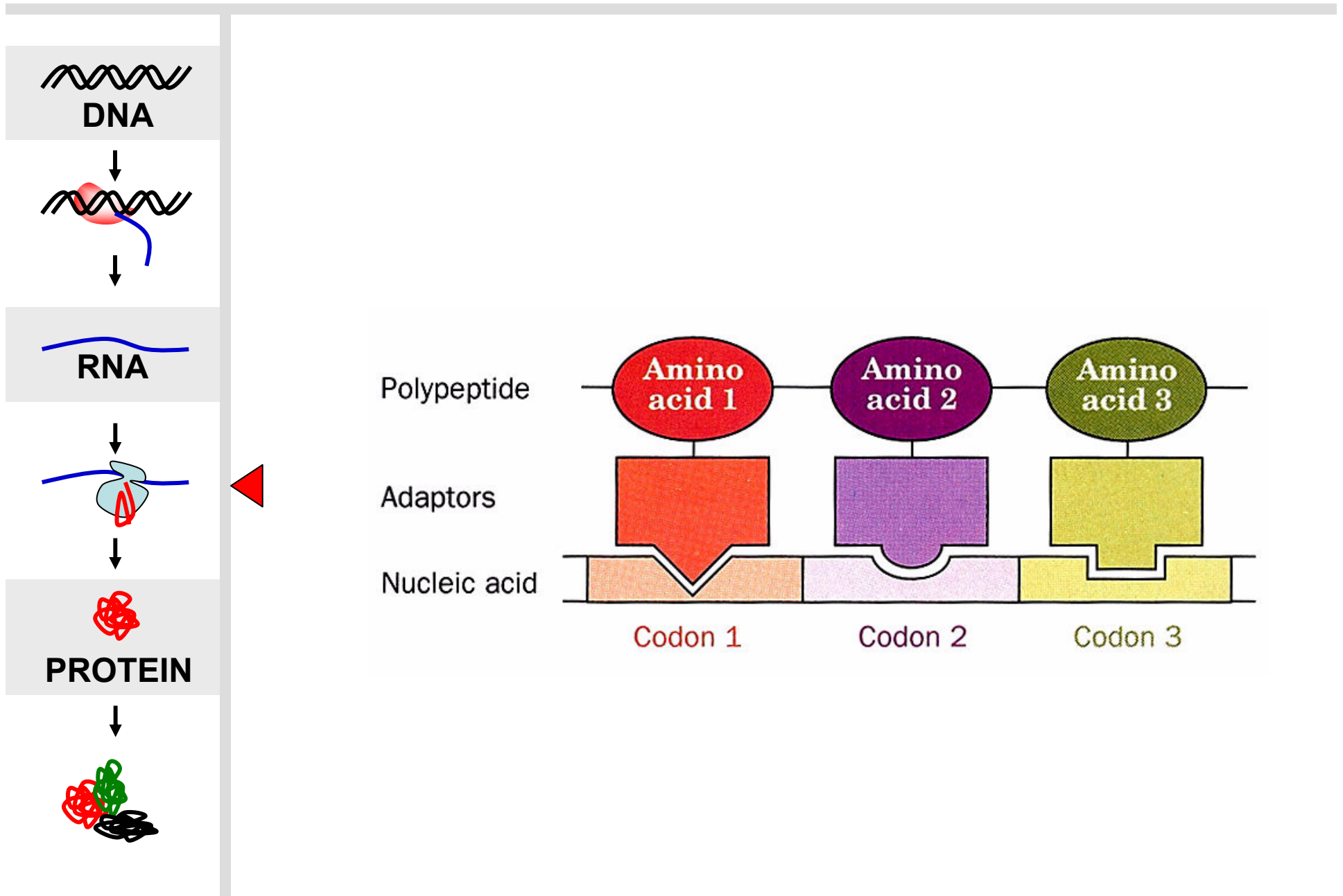
# Messenger RNA



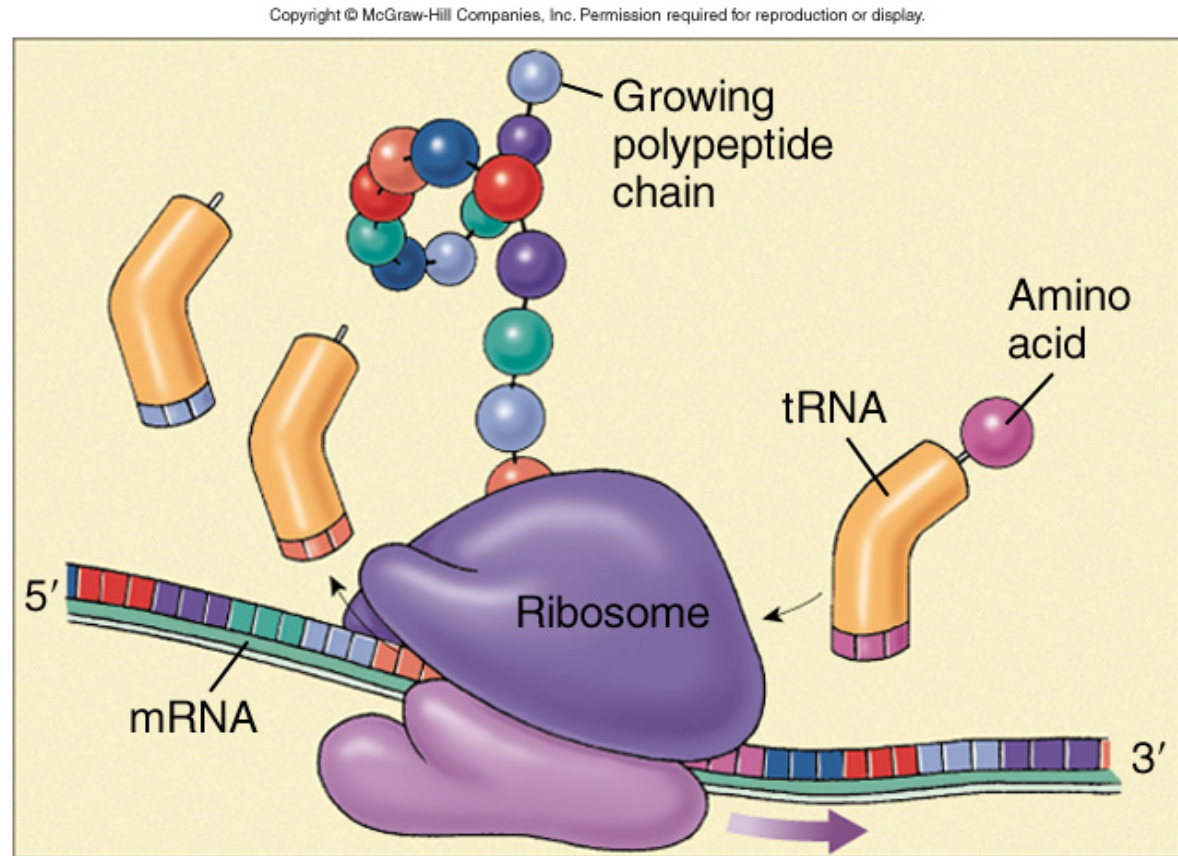
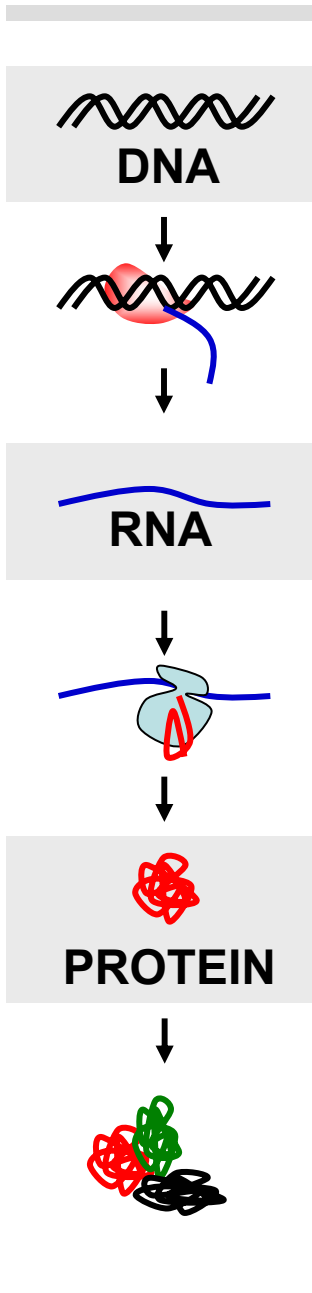
# Transcription



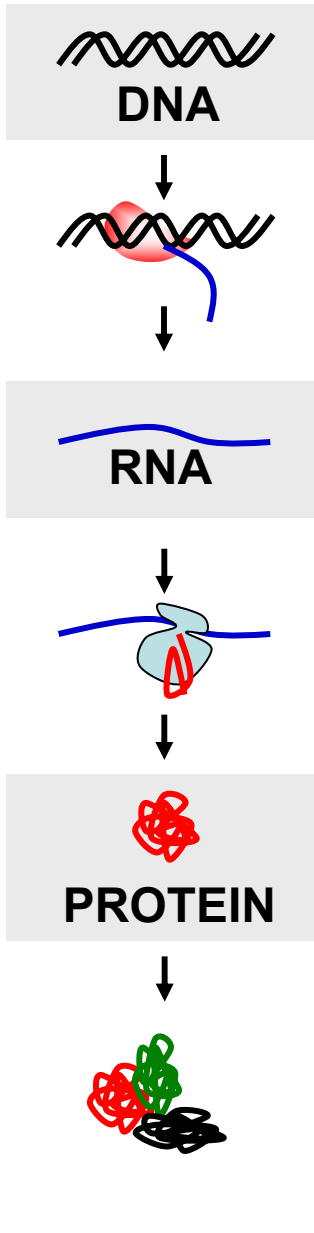
# Translation



# Translation



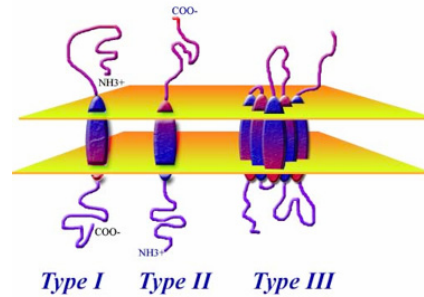
# Proteins and their Functions



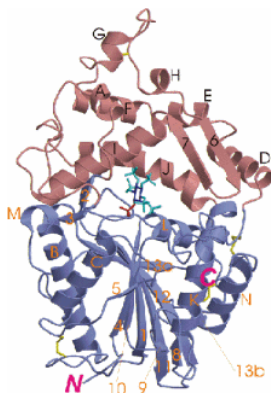
In the cell, proteins can:



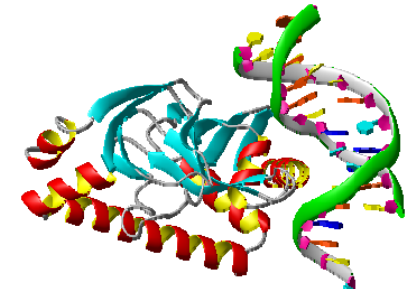
work as channels



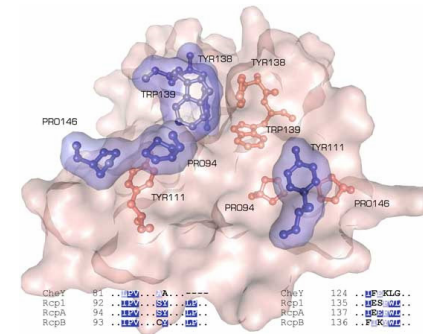
span the membrane for transport, signalling, ...



perform enzymatic reactions



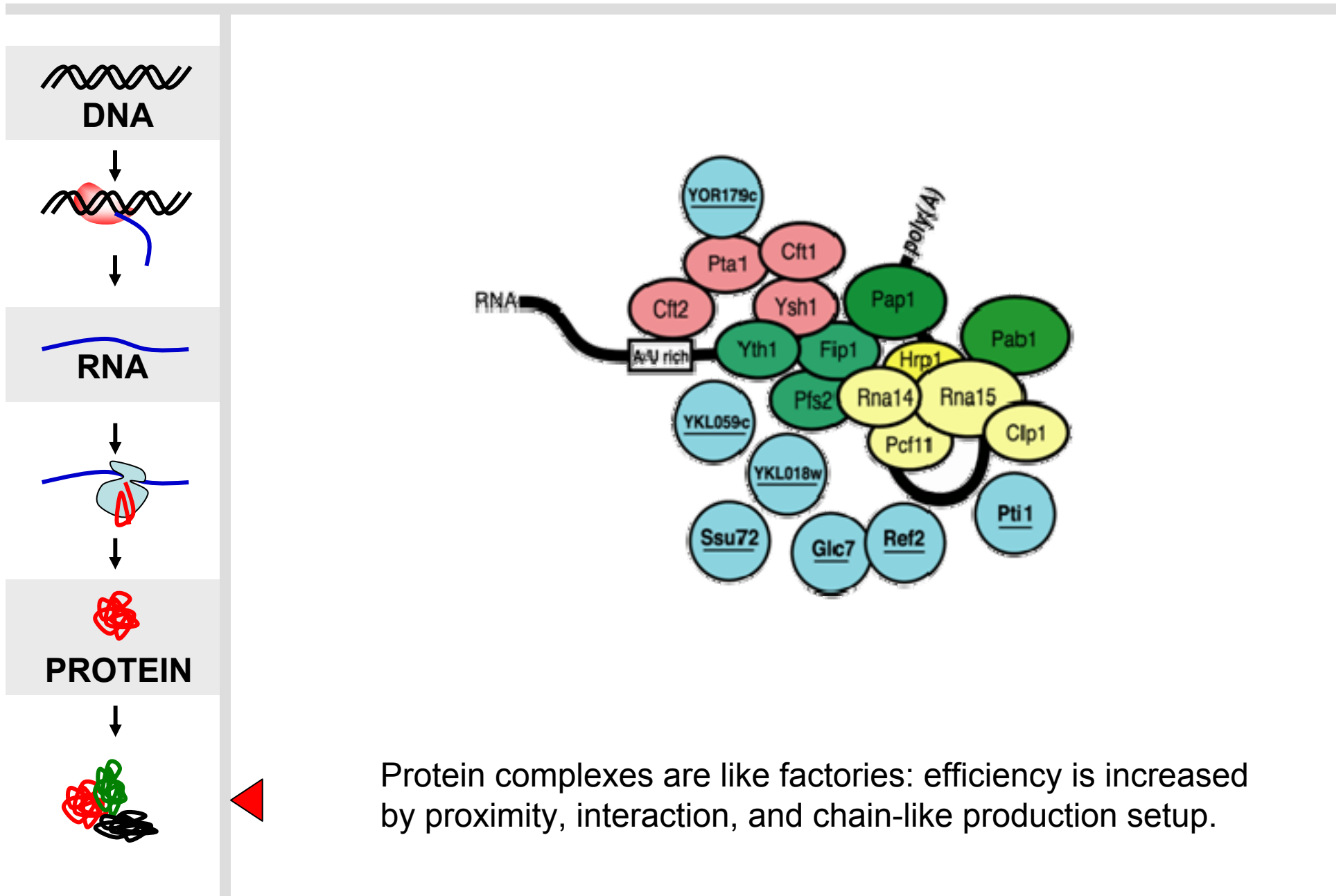
bind DNA



serve as ligands

... or can have many other functions

# Protein Complexes



Protein complexes are like factories: efficiency is increased by proximity, interaction, and chain-like production setup.



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high throughput technologies

# Types of Experiments

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## What to target.

- different developmental stages
- different organs

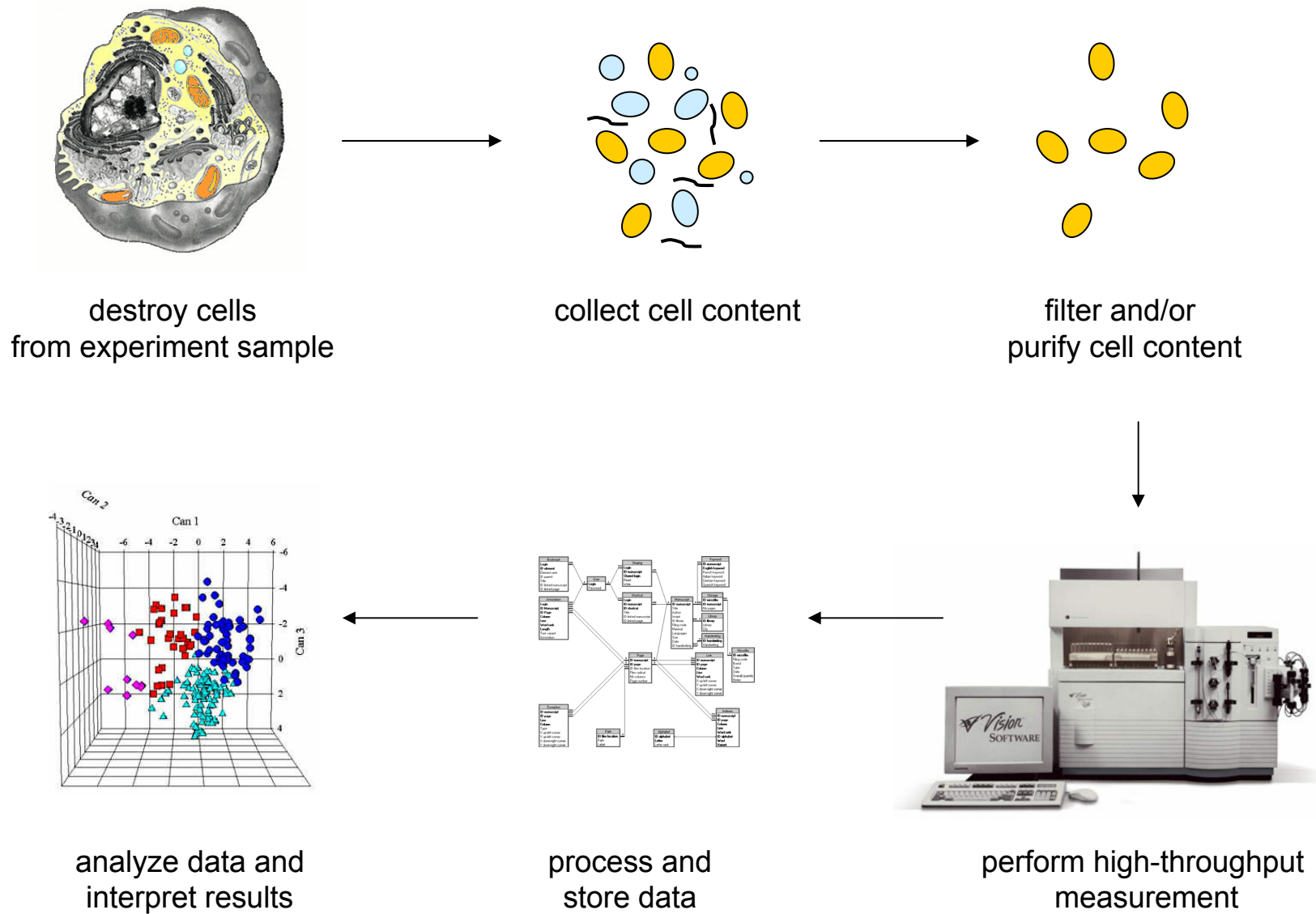
## How to perturb an organism.

- knock-out / knock-down
  - over-expressions
  - stimulus
  - disease
- } change expression of specific genes

## How to measure.

- steady state measurement
- time course measurements

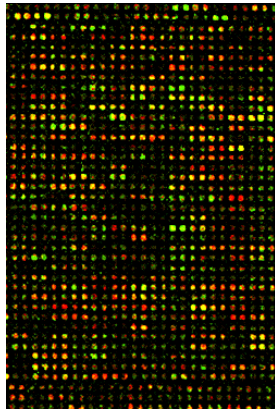
# Typical Workflow of an -omics Experiment



# DNA Microarrays

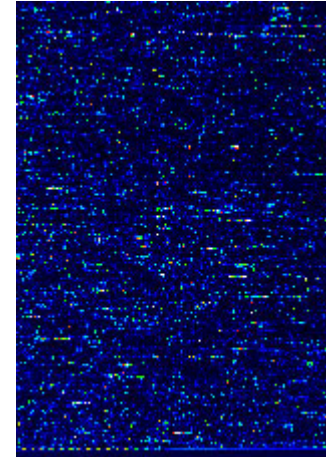
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## cDNA or GST Arrays



Dual dye (red/green)  
(Many companies)

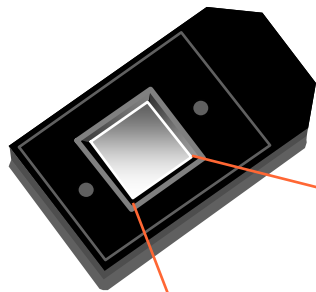
## Oligonucleotide Arrays



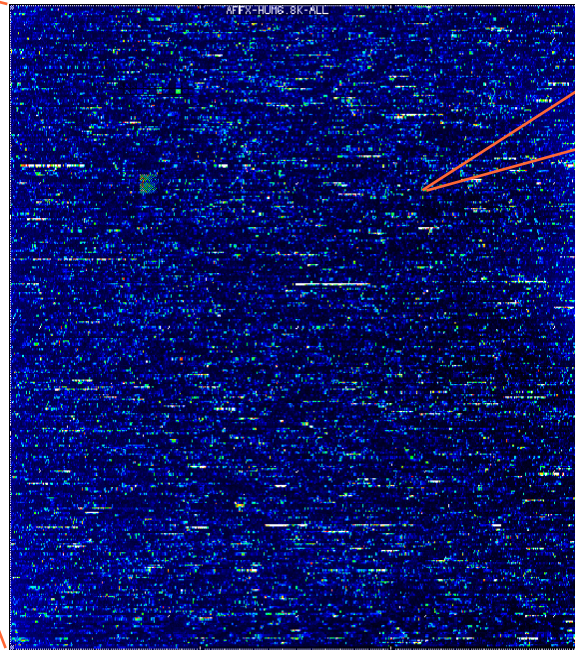
Single dye (Affymetrix, Nimblegen)  
Dual or single dye (Agilent)

# DNA Microarrays: Oligonucleotide Arrays

## GeneChip Probe Array

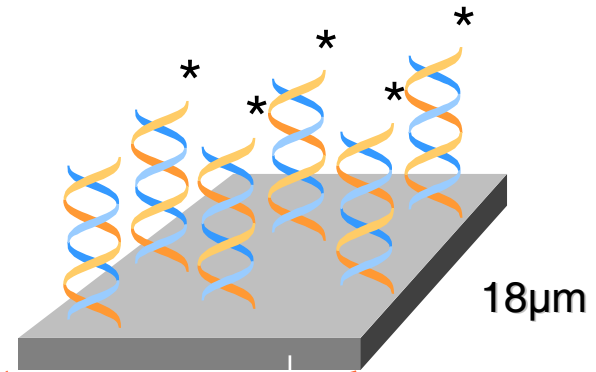


1.28cm



Scanned image

Single stranded,  
labeled RNA target  
Oligonucleotide probe



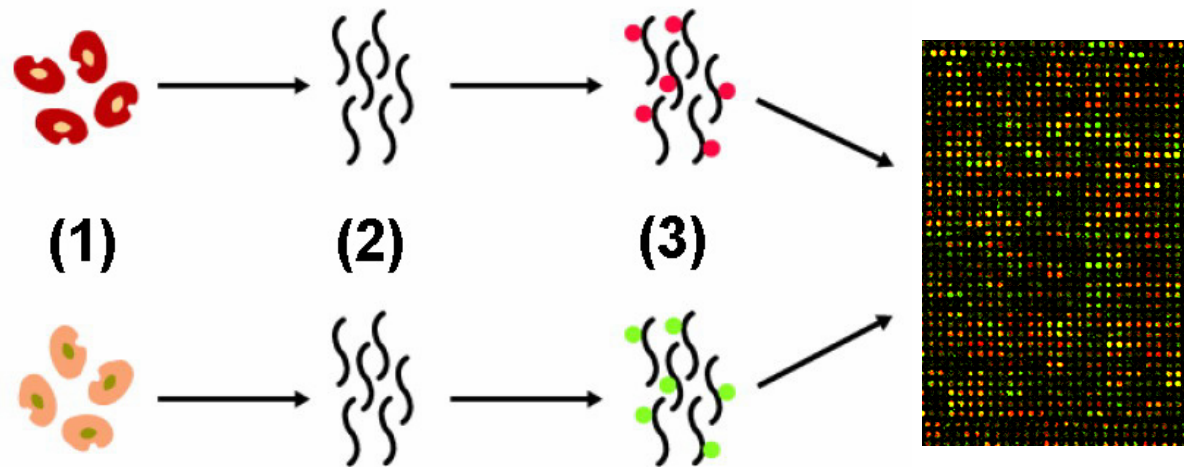
$10^6$ - $10^7$  copies of a specific  
oligonucleotide probe per feature

>1'000'000 different probes

# DNA microarrays: cDNA arrays

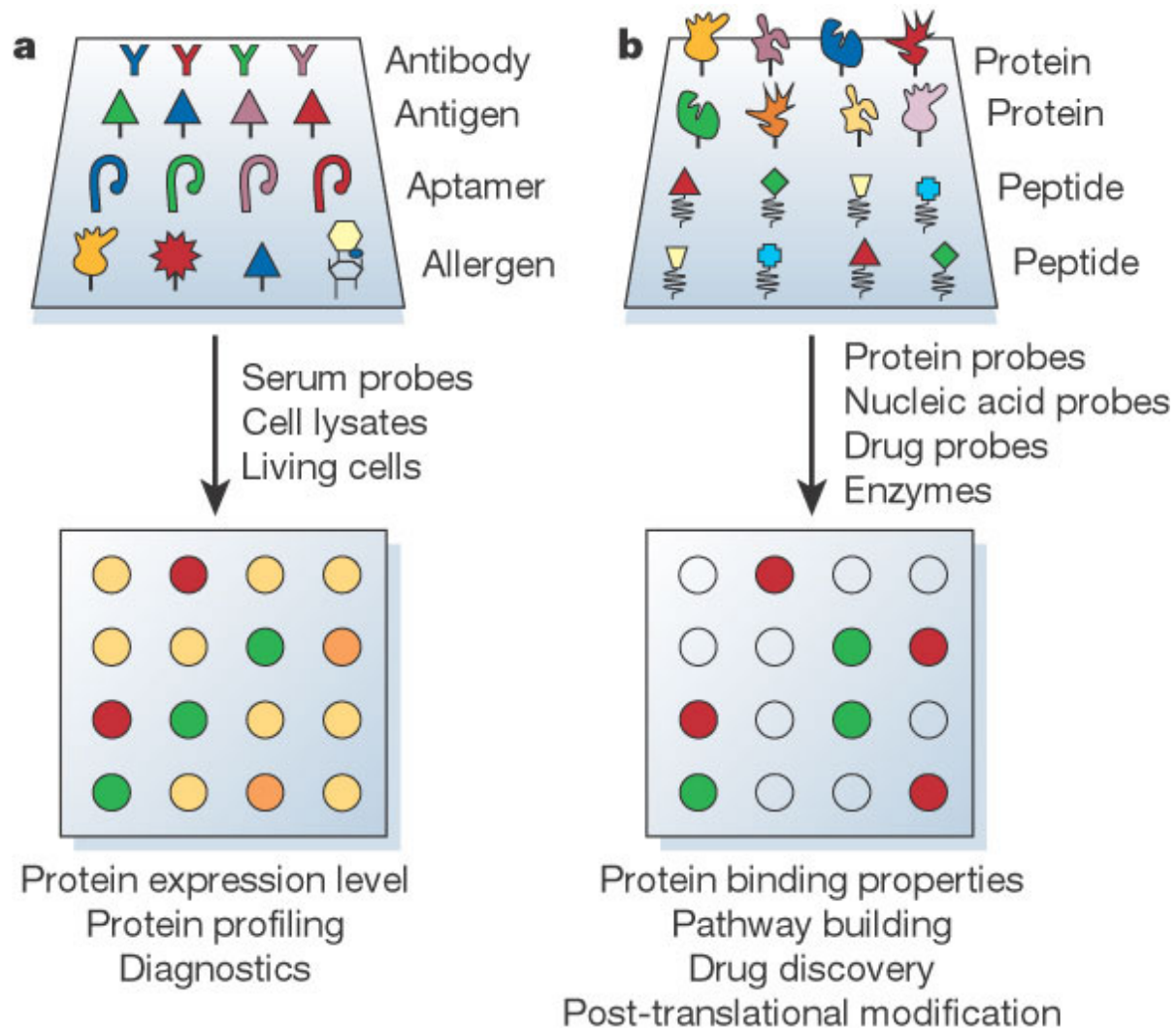
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Treatment



Control

# Protein Microarrays



# Proteomics

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Proteomics = study of the protein repertoire expressed in the cell

## Measurements

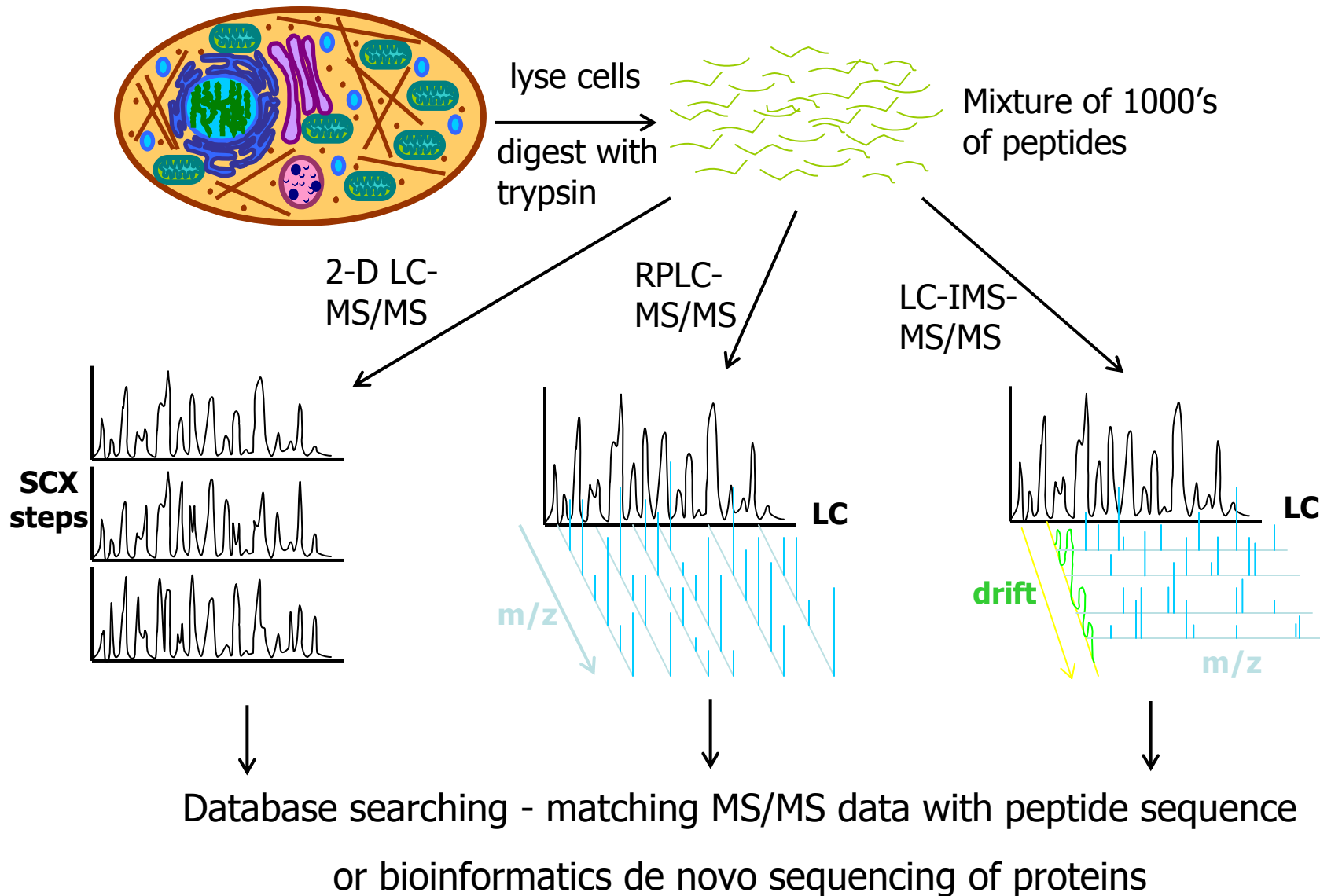
- protein expression levels (quantitative and qualitative)
- localization
- protein interaction

## Protein interactions elucidate...

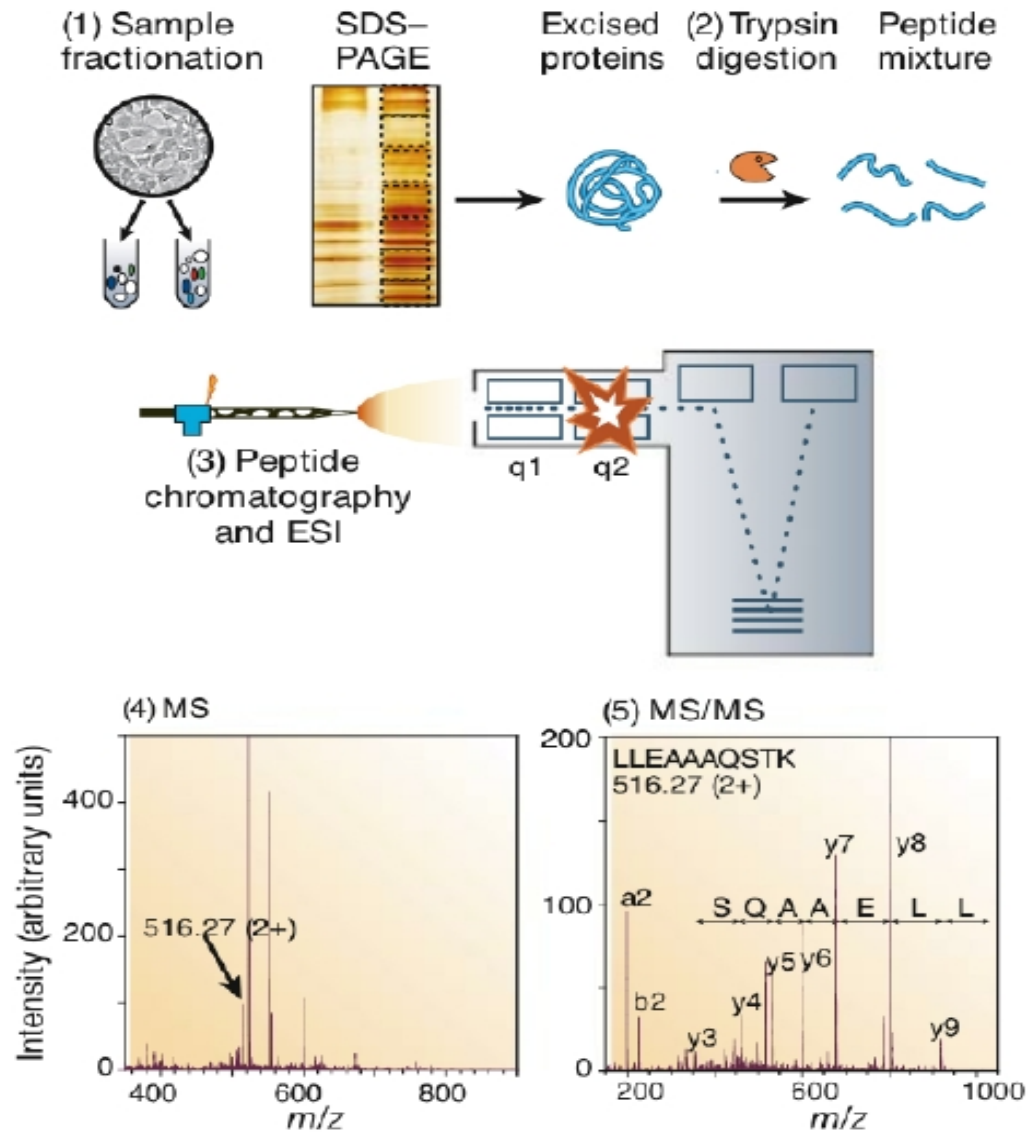
- pair-wise interactions
- protein complexes



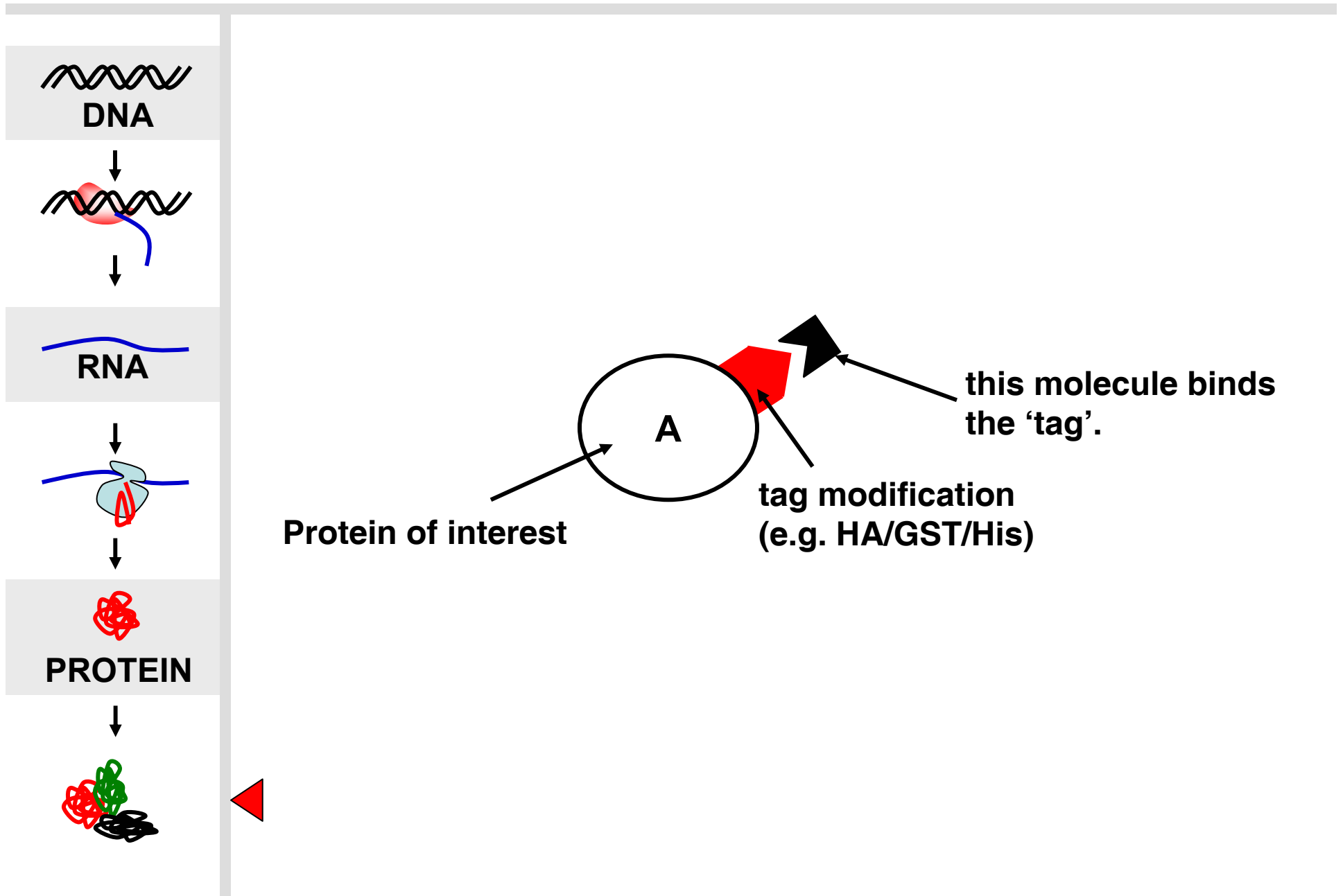
# Shotgun Proteomics



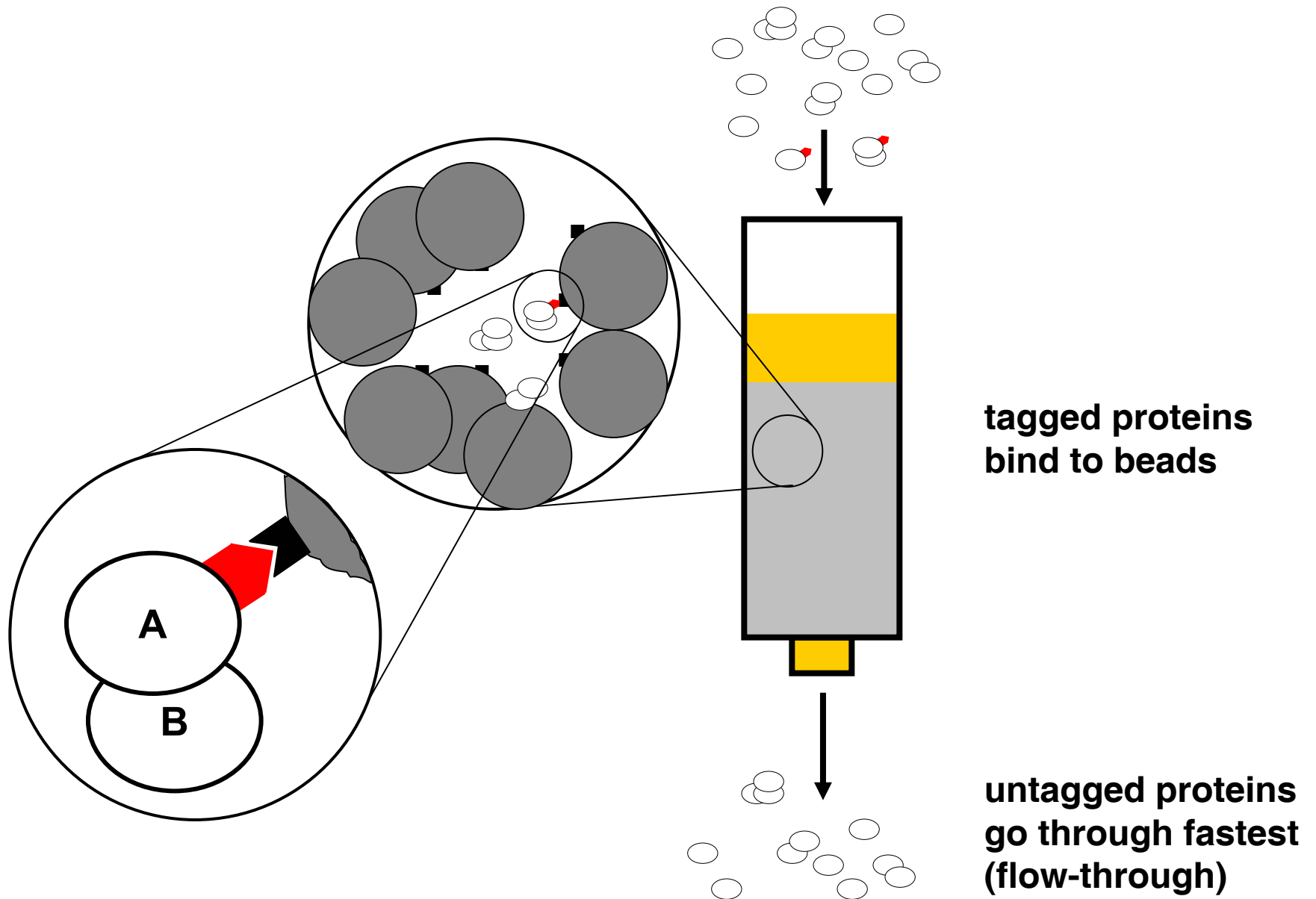
# Proteomics: Mass-Spectrometry Analysis



# Tandem Affinity Purification

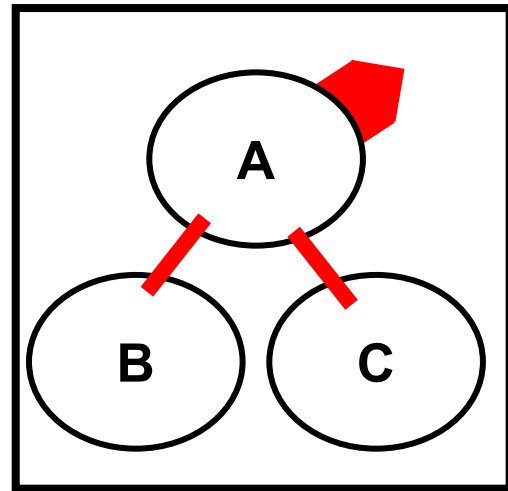
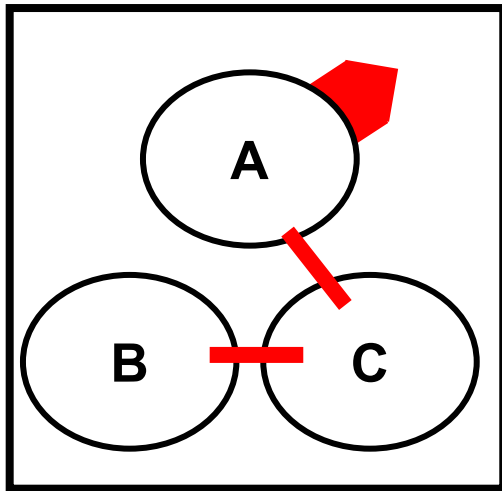
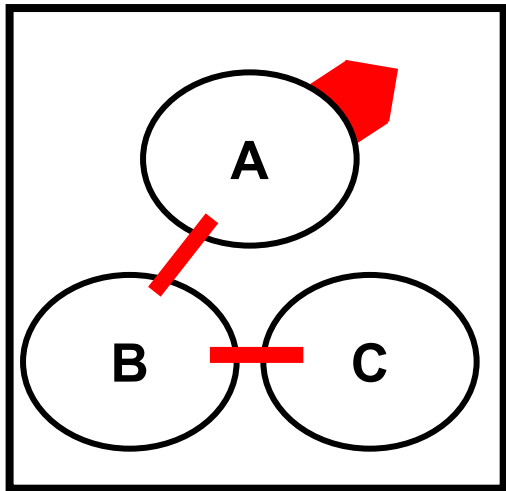


# Tandem Affinity Purification (TAP)

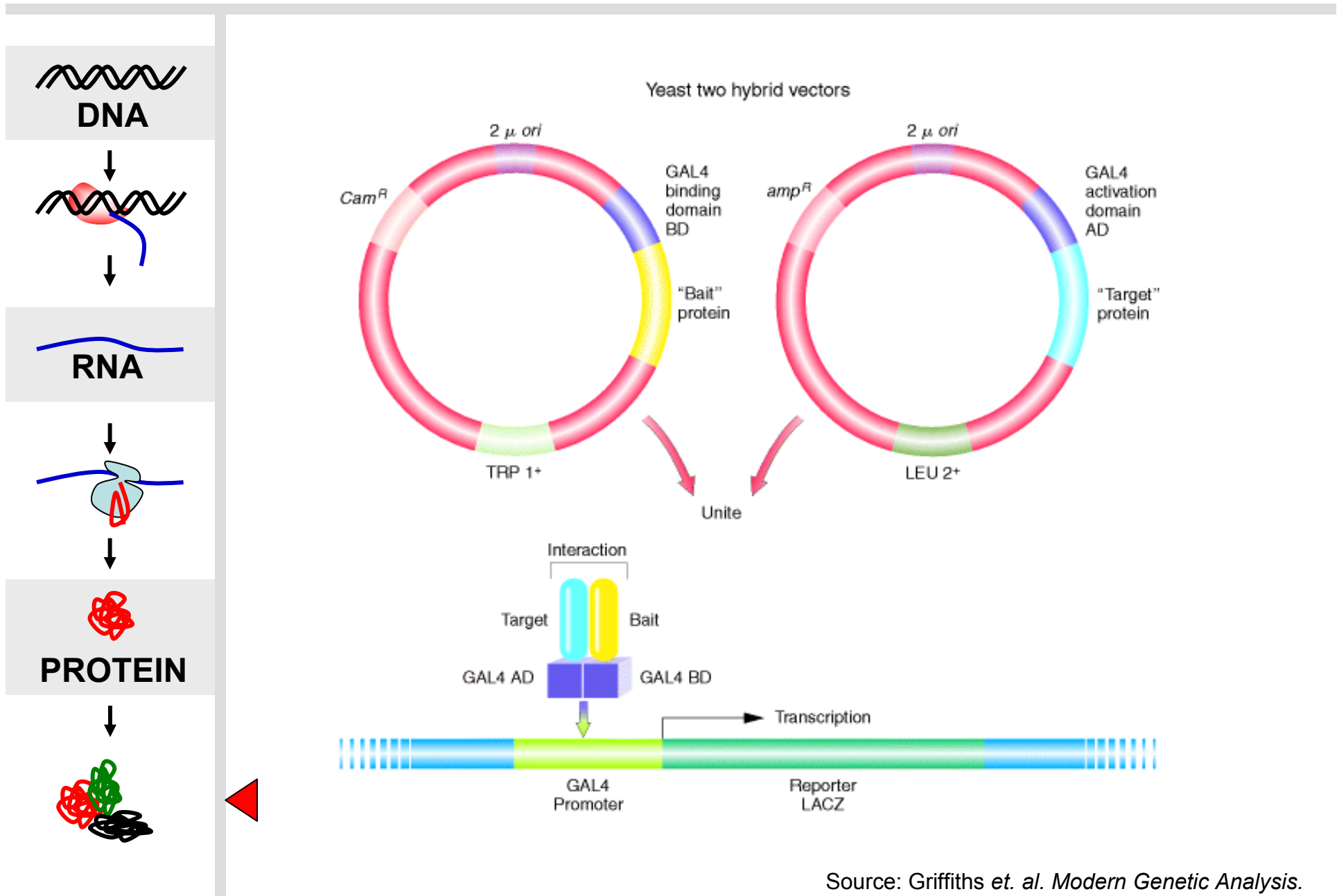


# TAP

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# Yeast two-hybrid

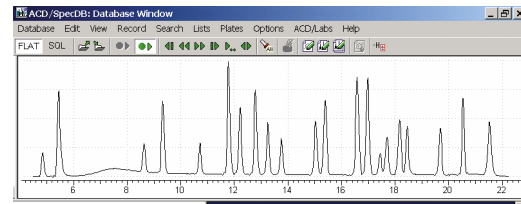


Source: Griffiths et. al. *Modern Genetic Analysis*.

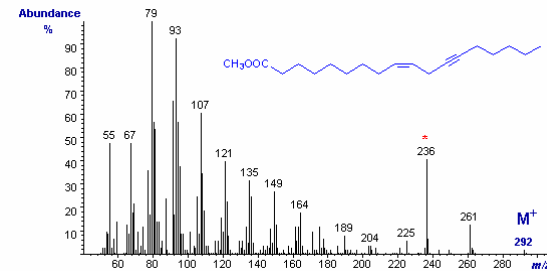
# Metabolomics

- Metabolomic methods:

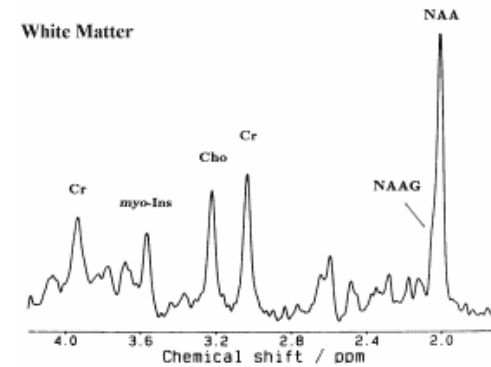
- Chromatography



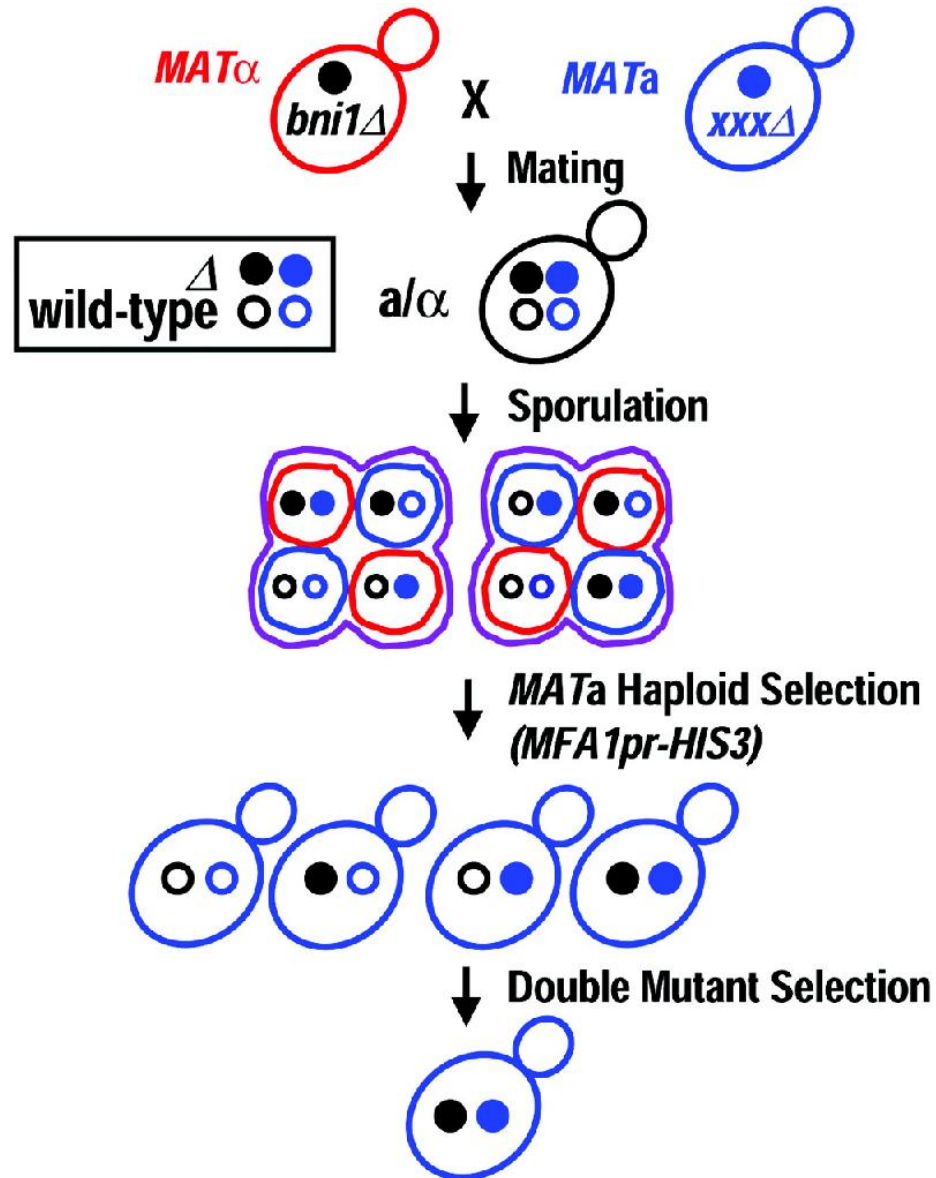
- Mass spectrometry (MS)



- Nuclear magnetic resonance (NMR)



# Synthetic Lethal Interactions

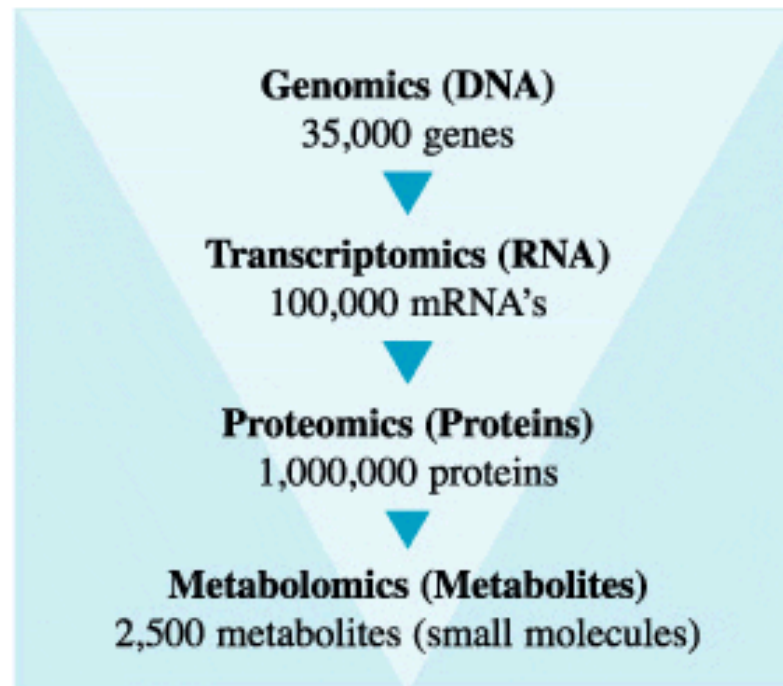




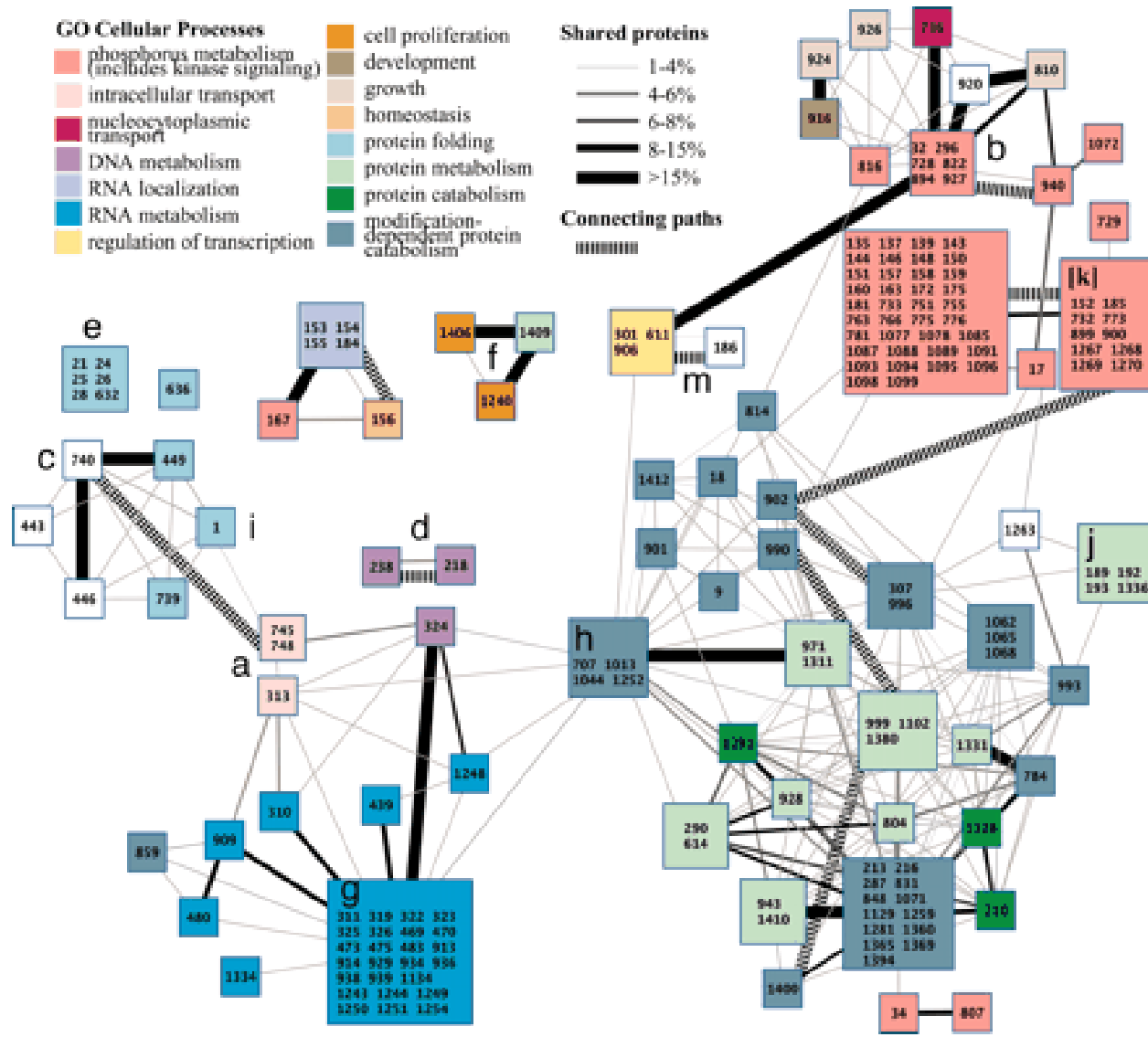
# OMICS...

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**Gene - Genome - Genomics**  
**Protein - Proteome - Proteomics**  
**Metabolite - Metabolome - Metabolomics**



# Complex Systems



# Formalized Biological Knowledge

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scientific literature

functional annotation

Gene Ontology  
swissprot

pathway databases

KEGG

phenotype and patient information

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## 2. Computational Issues in Systems Biology

# Computational Challenges

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experimental design

data preprocessing

data visualization

module identification

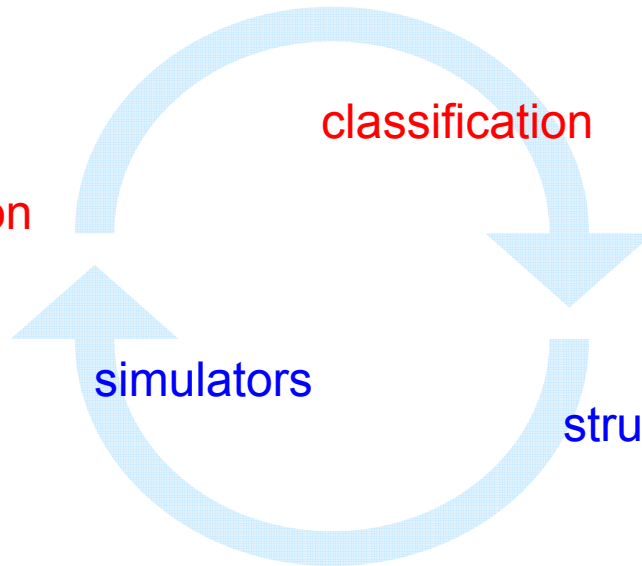
classification

simulators

structure prediction

network inference

text mining



# Classification of Tumor Samples – Problem

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## Goals

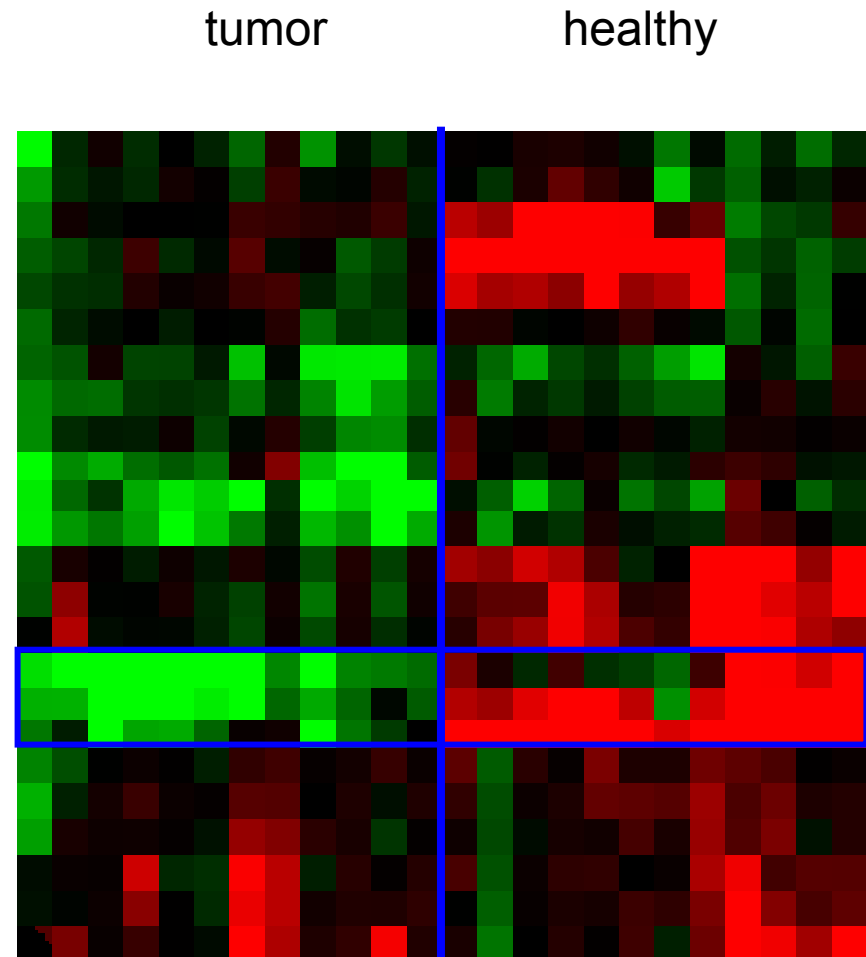
- discrimination between classes
- feature extraction

## Data

- mostly gene expression
- proteomics
- known outcome

## Challenges

- noisy data
- few samples, high dimensionality
- overfitting
- multiple testing



# Classification of Tumor Samples – General Approach

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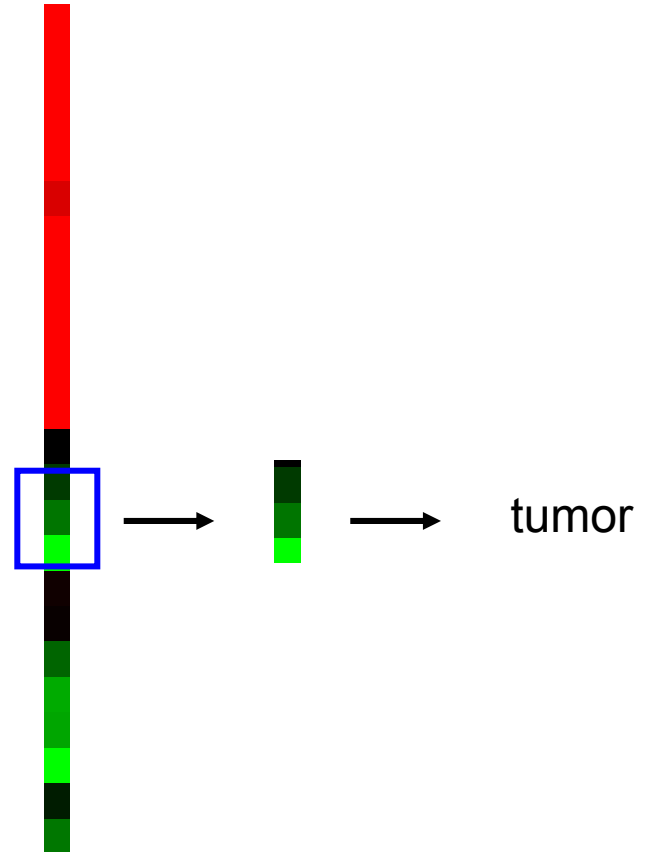
## Ingredients

- gene set selection
- classifier
- objective function
- optimizer

## Fighting Overfitting

- cross validation in objective function
- keep models small

new sample → class prediction



# Classification of Tumor Sample – EC approaches

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## Optimization Approaches

- genetic programming (GP) [1, 6, 7]
- simulated annealing [4]
- multiobjective evolutionary algorithm (including size) [3, 5]

- [1] J. Moore et al, **Symbolic Discriminant Analysis for Mining Gene Expression Patterns**, EMCL, 2001
- [2] J. Liu et al., **Selecting Informative Genes with Parallel Genetic Algorithms in Tissue Classification**, Genome Informatics, 2001
- [3] J. Liu et al., **Selecting Informative Genes Using a Multiobjective Evolutionary Algorithm**, WCCI, 2002
- [4] J. M. Deutsch, **Evolutionary algorithms for finding optimal gene sets in microarray prediction**, Bioinformatics, 2003
- [5] A. R. Reddy and K. Deb, **Identification of Multiple Gene Subsets Using Multi-objective Evolutionary Algorithms**, EMO, 2003
- [6] J. Rowland, **Model Selection Methodology in Supervised Learning with Evolutionary Computation**, Biosystems, 2003
- [7] W. B. Langdon and B. F. Buxton, **Genetic Programming for Mining DNA Chip data from Cancer Patients**, Genetic Programming and Evolvable Machines, 2004
- [8] J. Rowland, **On Genetic Programming and Knowledge Discovery in Transcriptome Data**, CEC, 2004



# Classification of Tumor Samples - Moore et al. [1]

## Individual

- real valued function  $f$  of gene expression
- represented as GP tree

## Classifier

- $f >$  median of all function values

## Objective Function

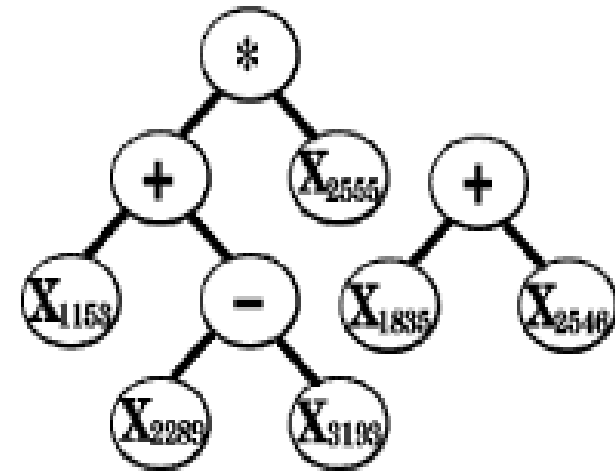
- classification error

## Optimizer

- parallel GP

## Results

- data: AML/ALL (Golub et al.) two class problem
- two nearly perfect predictors:
  - $X_{1835} + X_{2546}$
  - $X_{2555} * (X_{1153} + X_{2289} + X_{3193})$



# Classification of Tumor Samples - Deutsch [4]

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## Individual

- set of predictive genes
- represented as list

## Classifier

- k-nearest neighbor ( $k = 1$ )

## Objective Function

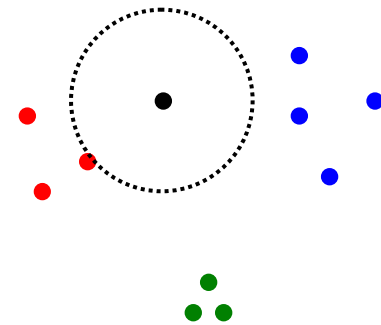
- weighted sum of LOOCV accuracy and clustering score

## Optimizer

- variant of simulated annealing (replication algorithm)
- mutation: add or remove one gene

## Results

- data: multiple data sets (incl. one with more than 2 classes)
- results: smaller gene sets and good classification



# Classification of Tumor Samples – Liu et al. [3]

## Individual

- set of predictive genes
- represented as bit string

## Classifier

- normalized distance to class mean

## Objective Function

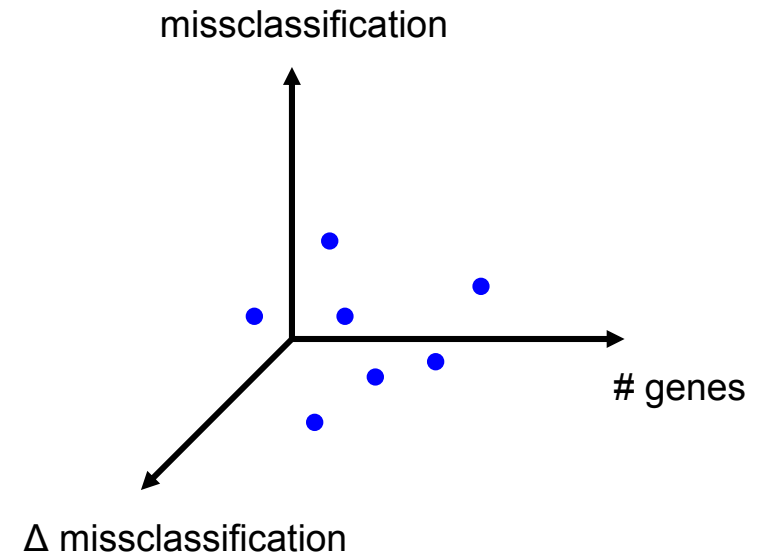
1. gene set size
2. missclassification rate
3. difference of missclassification rates

## Optimizer

- multiobjective EA
- called replication algorithm

## Results

- data: Leukemia, Lymphoma and Colon cancer data sets
- results: many diverse and small gene sets



# Classification of Tumor Samples – Langdon et al. [7]

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## Individual

- sum  $S$  of 5 real valued function of expression values
- represented as 5 GP trees

## Classifier

- $S > 0$

## Objective Function

- mean of LOOCV accuracies for both classes

## Optimizer

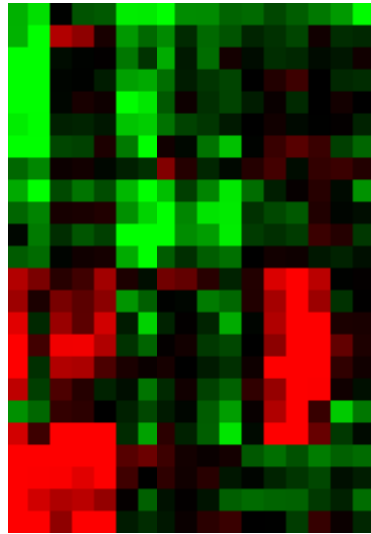
- GP

## Results

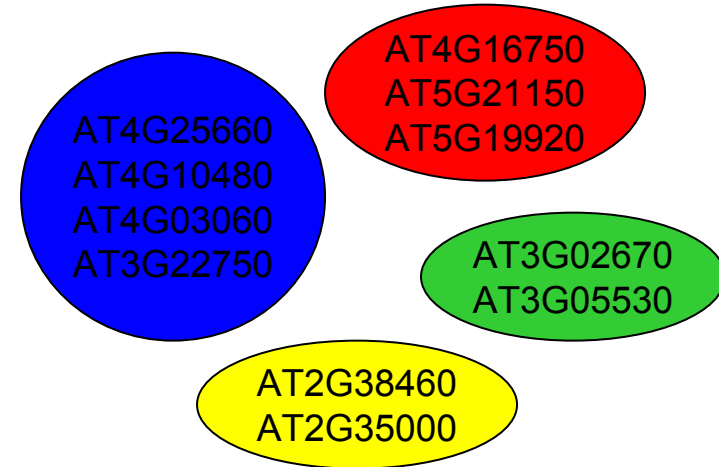
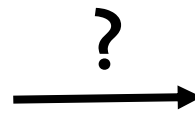
- data: Central nervous system embryonal tumors [Pomeroy et al. 2002]
- results: good classification, surprisingly small gene sets

# Module Identification

## Goal



high throughput data



functional gene groups

## Approaches

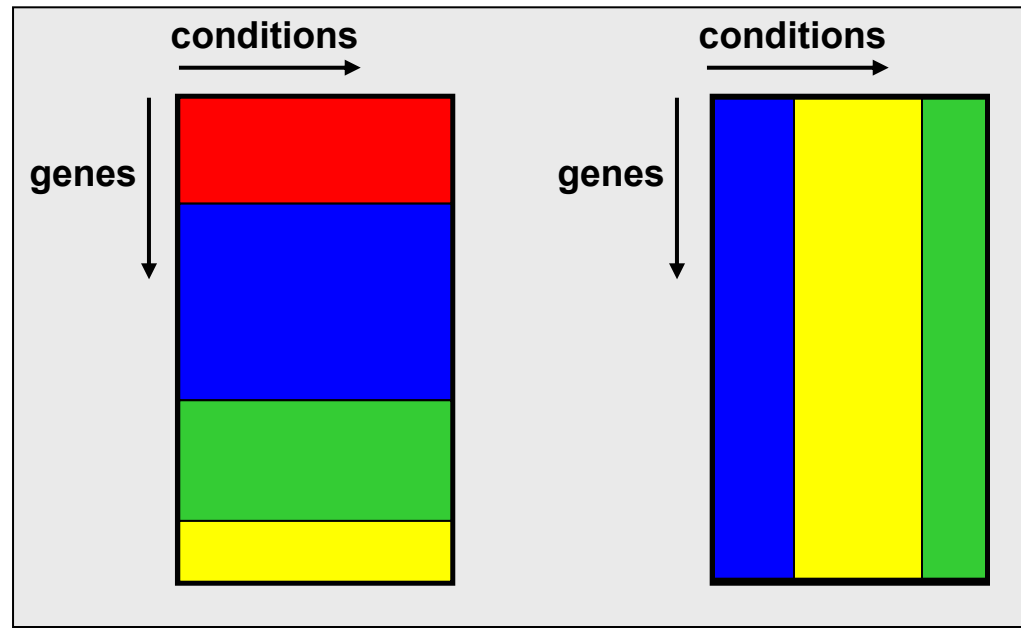
- guilt by association
- clustering, biclustering
- integration with additional data, e.g., promoter elements

## Challenges

- huge search space
- data integration

# Clustering of Gene Expression Data

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## Inputs

- gene expression data
- number of clusters

## Clustering algorithms ...

- ... group similar things.
- ... partition the matrix.
- ... use all measurements.

# Clustering with EC – Falkenauer et al. [9]

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## Individual

- clustering = partitioning of input matrix
- specific representation

## Objective Function

- total variance within clusters

## Optimizer

- Grouping Genetic Algorithm

## Results

- data: 3 different gene expression data sets
- results: much better than k-means (which uses the same objective function)

[9] E. Falkenauer and A. Marchand, **Clustering Microarray Data Using Evolutionary Algorithms**, chapter in “Evolutionary Computation in Bioinformatics”, Morgan Kaufmann, 2003

# Clustering with EC – Handl et al. [10]

## Individual

- clustering = partitioning of input matrix
- locus based adjacency representation

## Objective Function

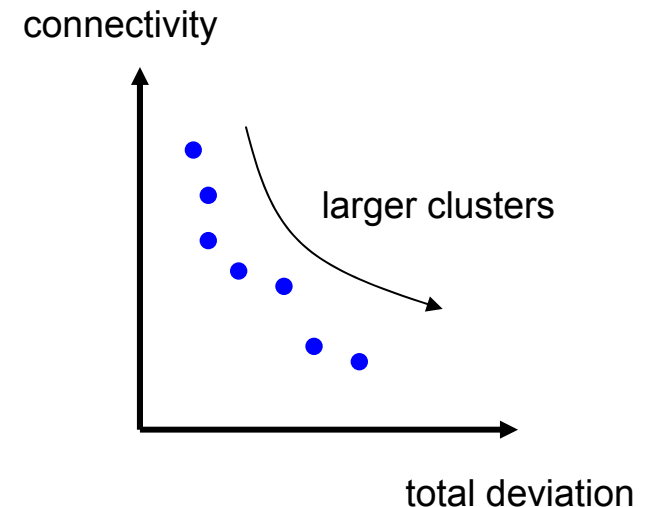
1. total deviation from cluster means
2. total connectivity (high if neighbors are not in the same cluster)

## Optimizer

- multiobjective EA (PESA-II)

## Results

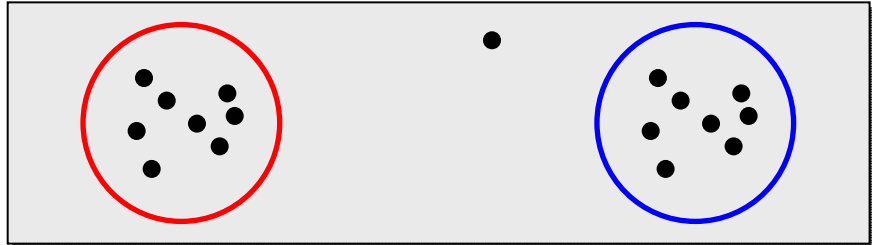
- good performance compared to k-means and average-linkage hierarchical clustering algorithms
- automatic determination of the number of clusters



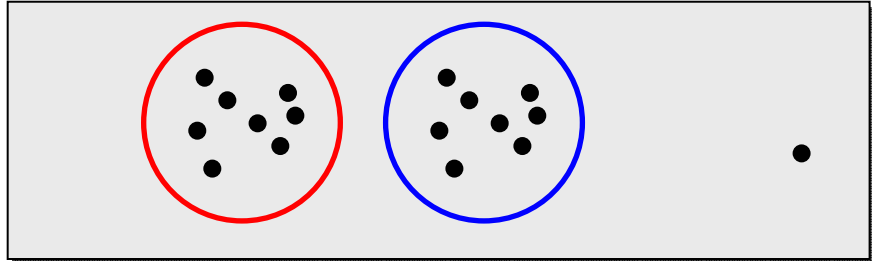


# Drawbacks of Standard Clustering

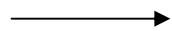
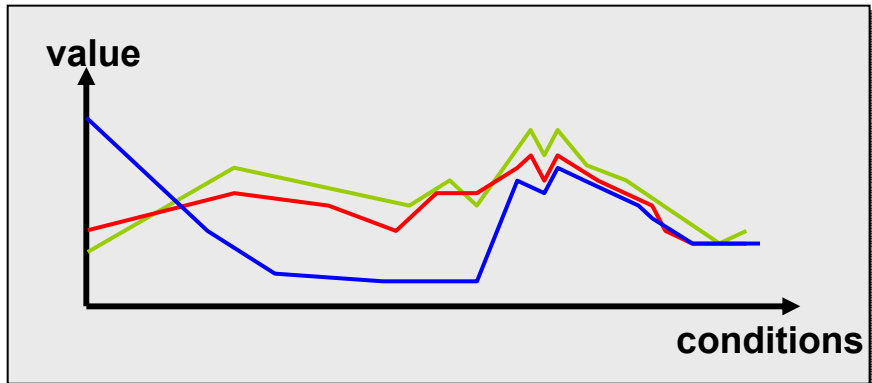
- a gene cannot be in two clusters



- each gene is assigned to a cluster



- local patterns are missed



**New problem formulation needed.**

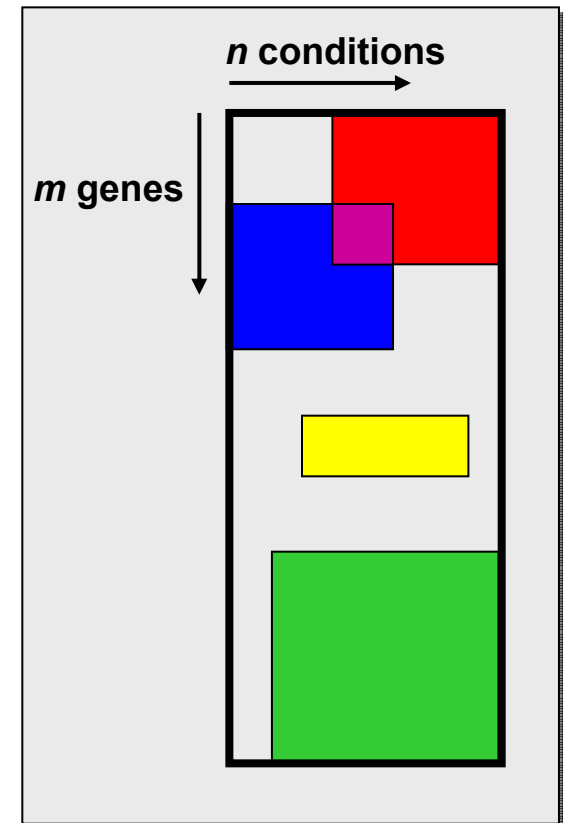
# Biclustering

## Goal

- find subsets of genes – subsets of conditions
- may overlap
- two objectives: size and similarity

## Existing Algorithms

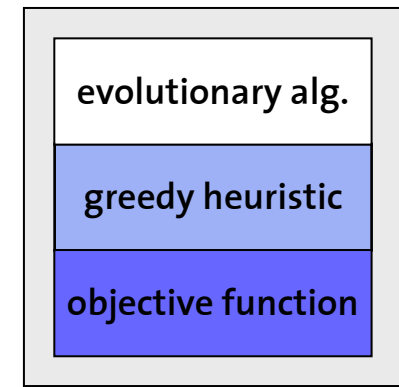
- definition of similarity
- number of biclusters
- search strategy



# EC for Biclustering – Bleuler et al. [11]

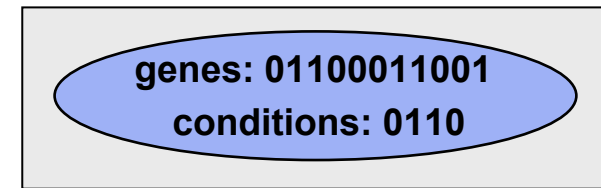
## Approach

- existing algorithms as local search
- EA as global search
- systematic sampling of the search space
- applicable to many existing algorithms



## Basics

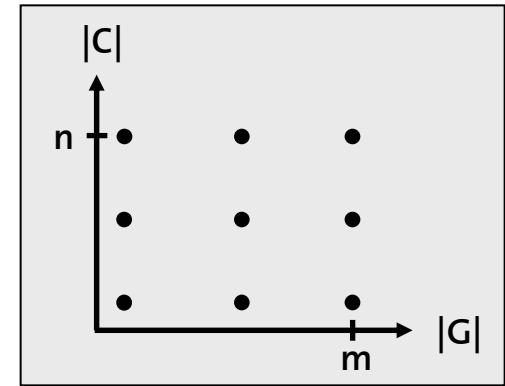
- individual = submatrix
- binary encoding
- independent bit mutation
- uniform crossover
- local search
- tournament selection ( $t \in \{3, 20\}$ )
- 100 individuals, 50 generations



# EC for Biclustering – Bleuler et al. [11]

## Initialization

- set each bit to 1 with  $p = 0.5$   
→ **biclusters have similar size**
- better: distribute bicluster sizes

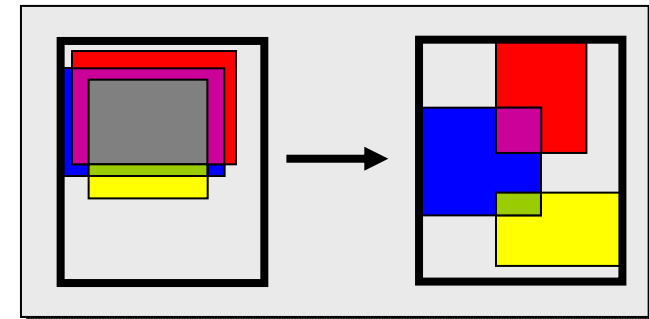


## After the local search...

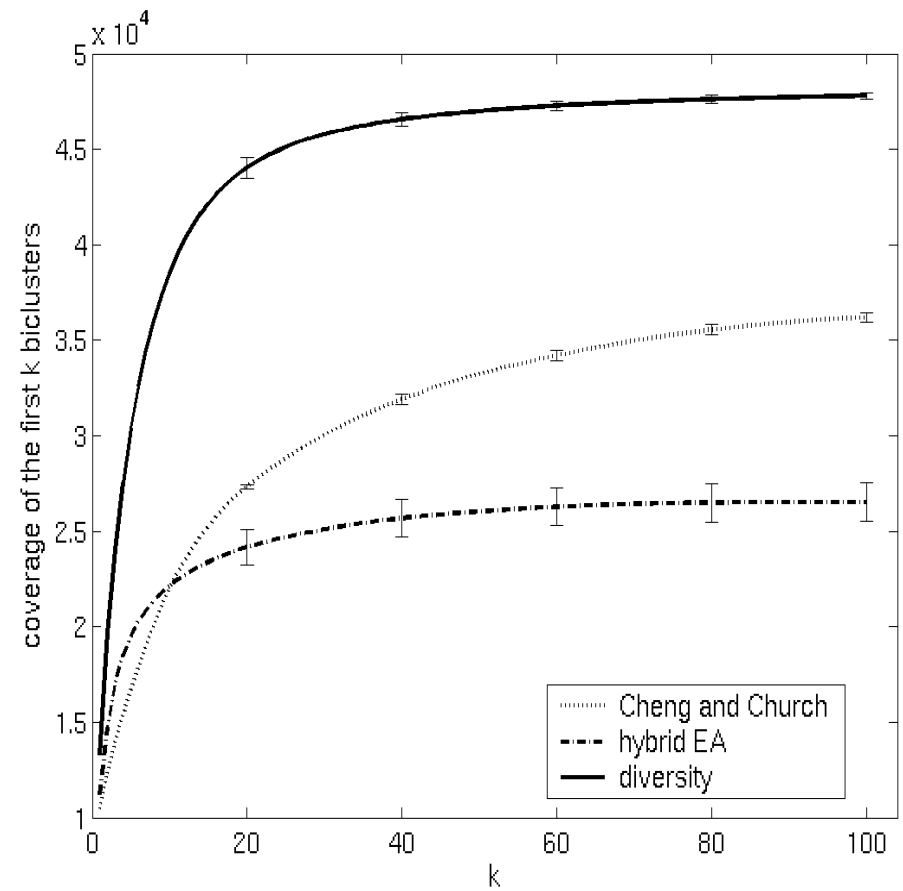
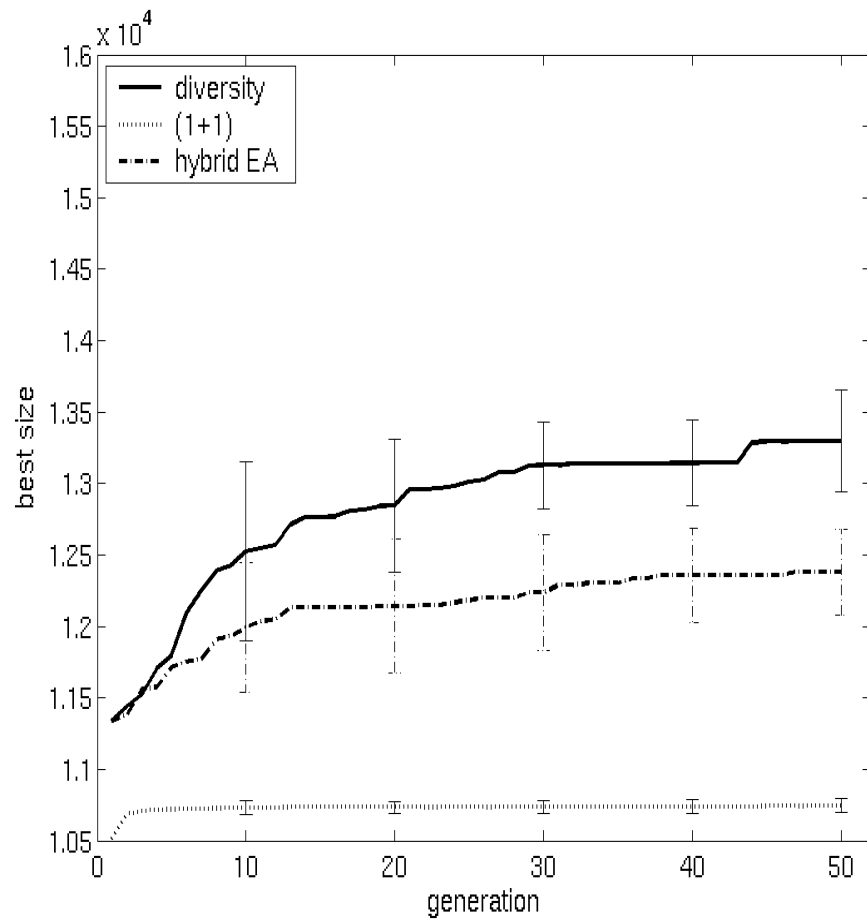
- update (Lamarckian evolution)
- don't update (Baldwinian evolution)

## Diversity Maintenance

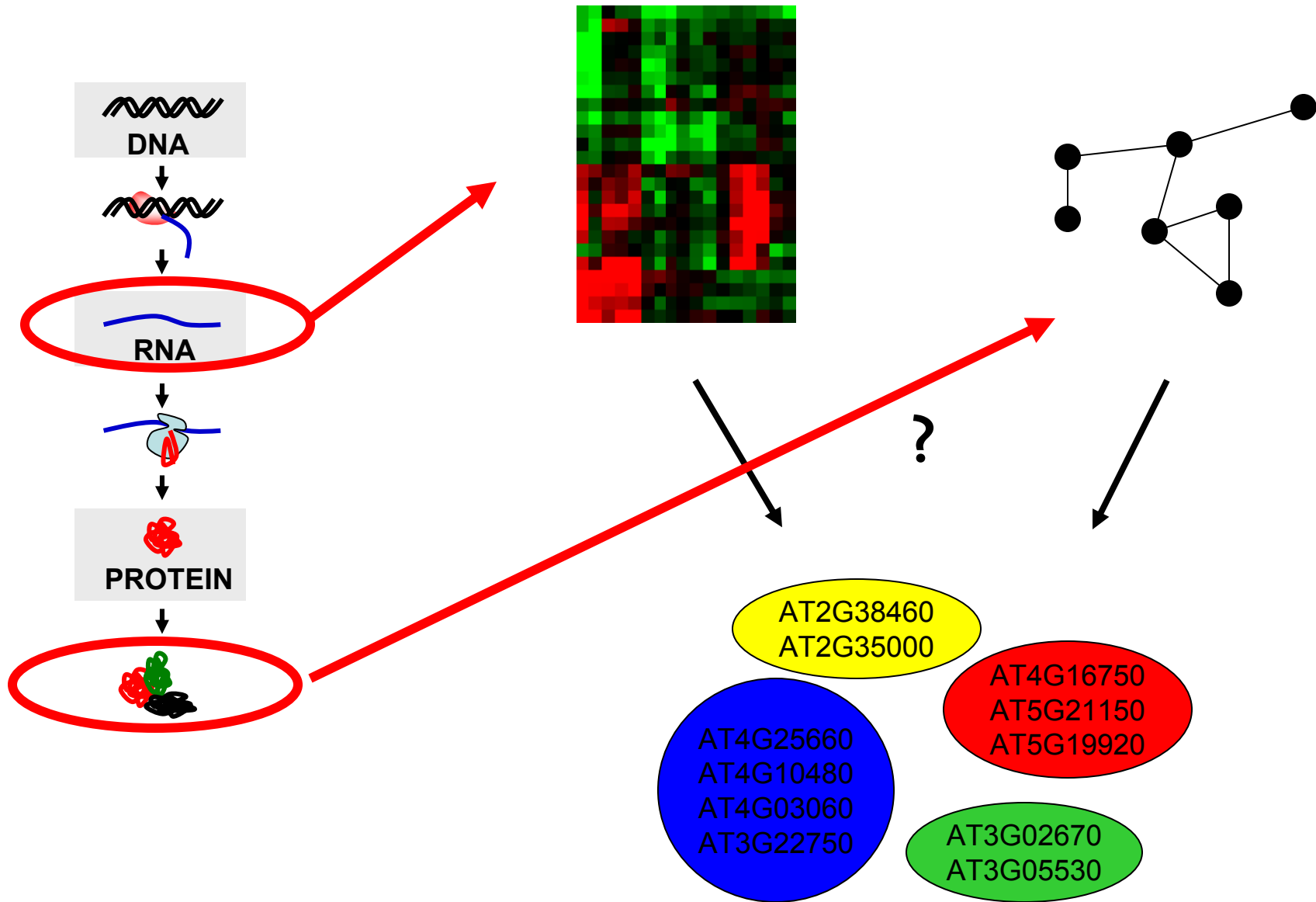
- N biclusters in one run
- optimize coverage
- select individual with most new area



# EC for Biclustering – Bleuler et al. [11]



# Gene Expression is not Enough!



# EC for Data Integration - Speer et al. [12]

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## Goal

- clustering of gene expression data and Gene Ontology graph

## Individual

- clustering = partitioning of input matrix
- representation based on minimum spanning tree
- represented as  $n-1$  bits determining whether to cut the MST at edge  $i$ .

## Objective Function

- weighted sum of distance on gene expression and distance on Gene Ontology graph

## Optimizer

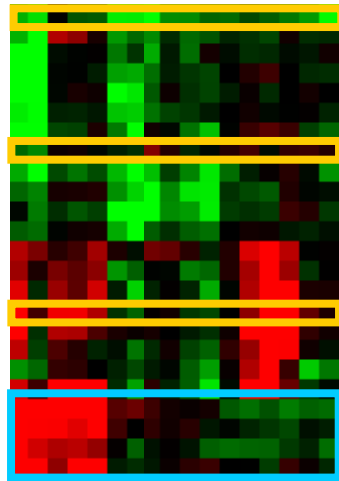
- EA with local search

## Results

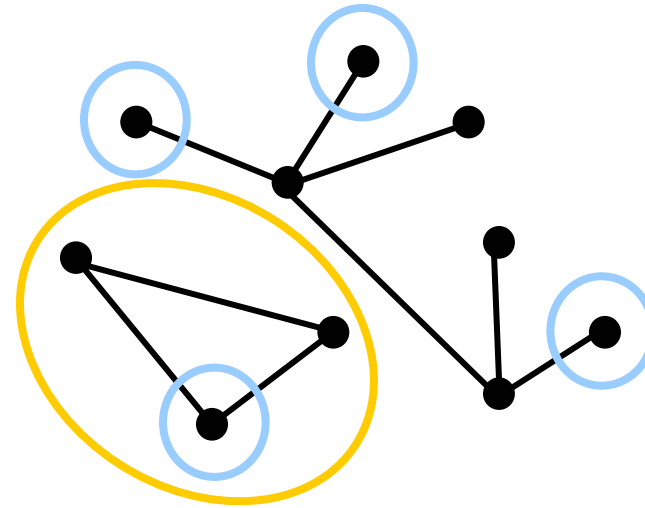
- data: gene expression data from human fibroblast and GO
- results: some clusters more gene expression oriented others more GO oriented

# EC for Data Integration - Bleuler et al. (work in progress)

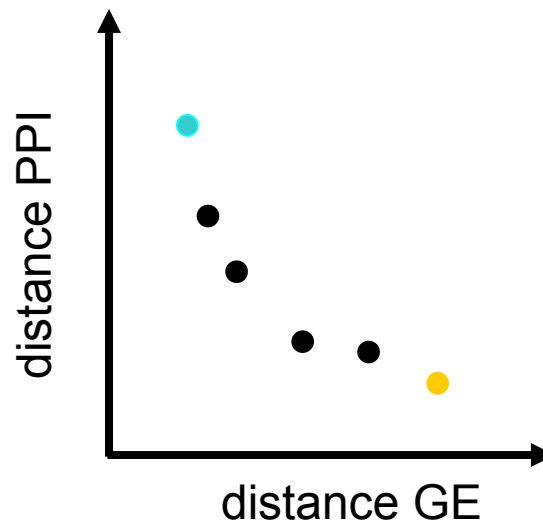
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gene expression

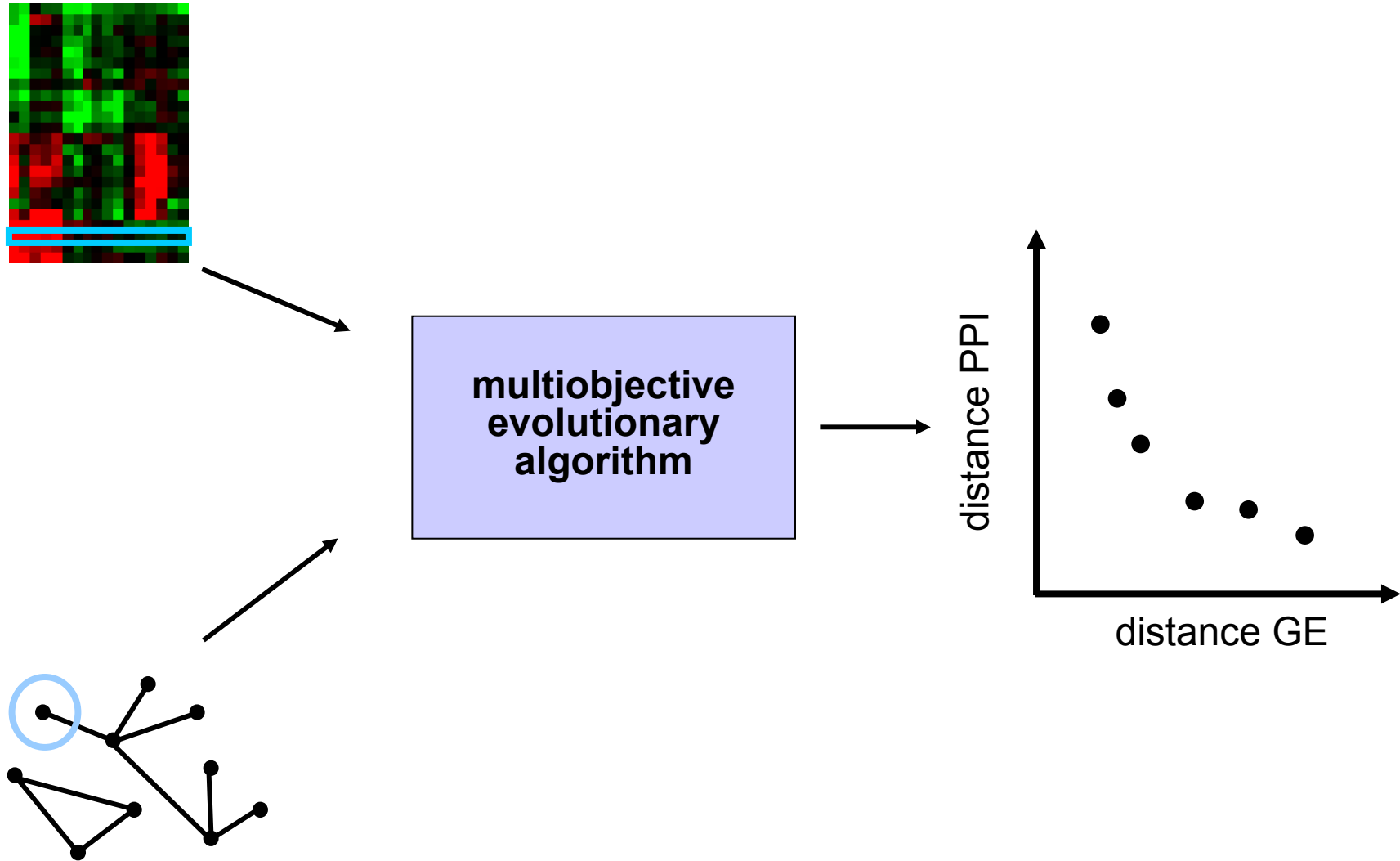


protein-protein interactions

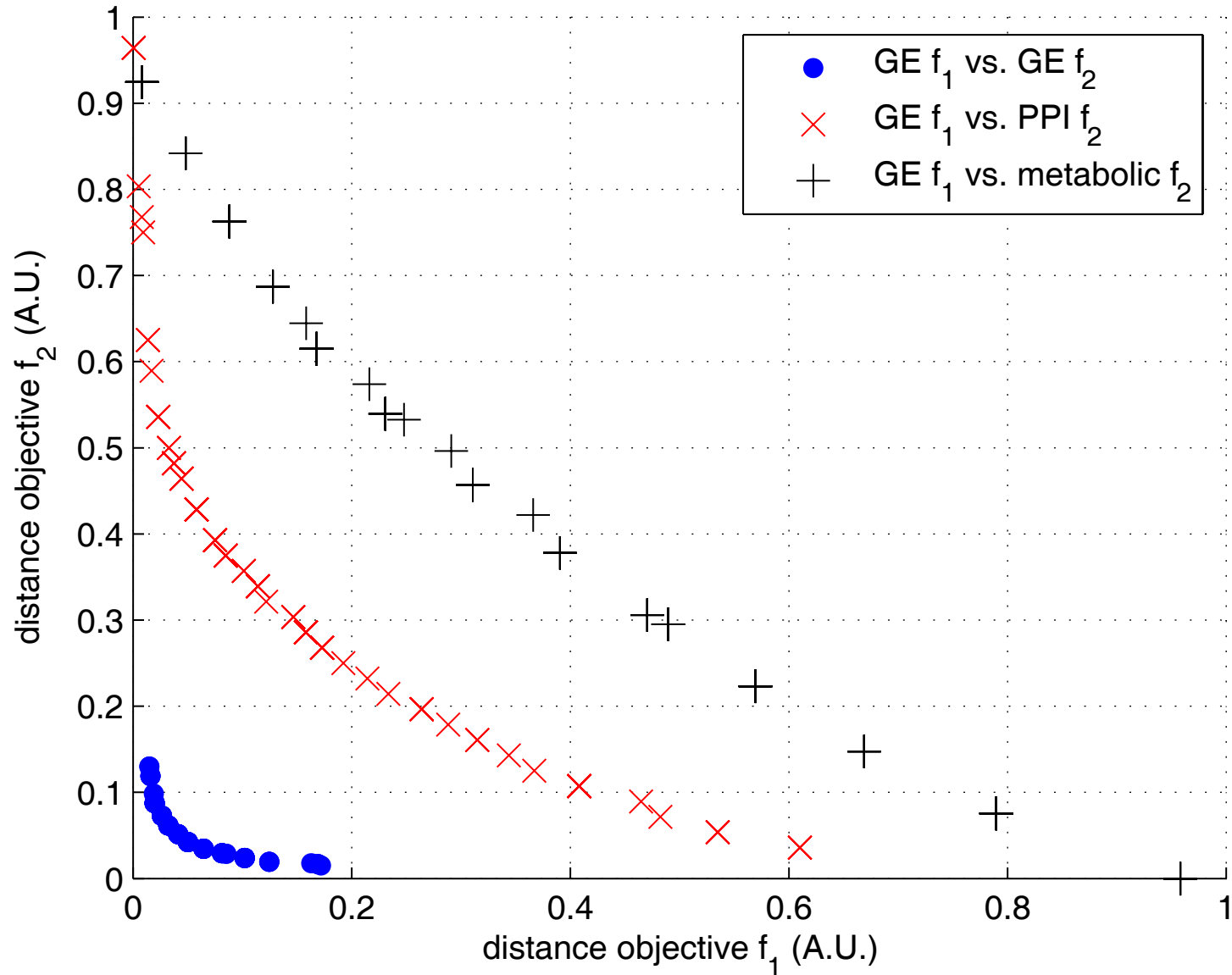




# EC for Data Integration - Bleuler et al. (work in progress)

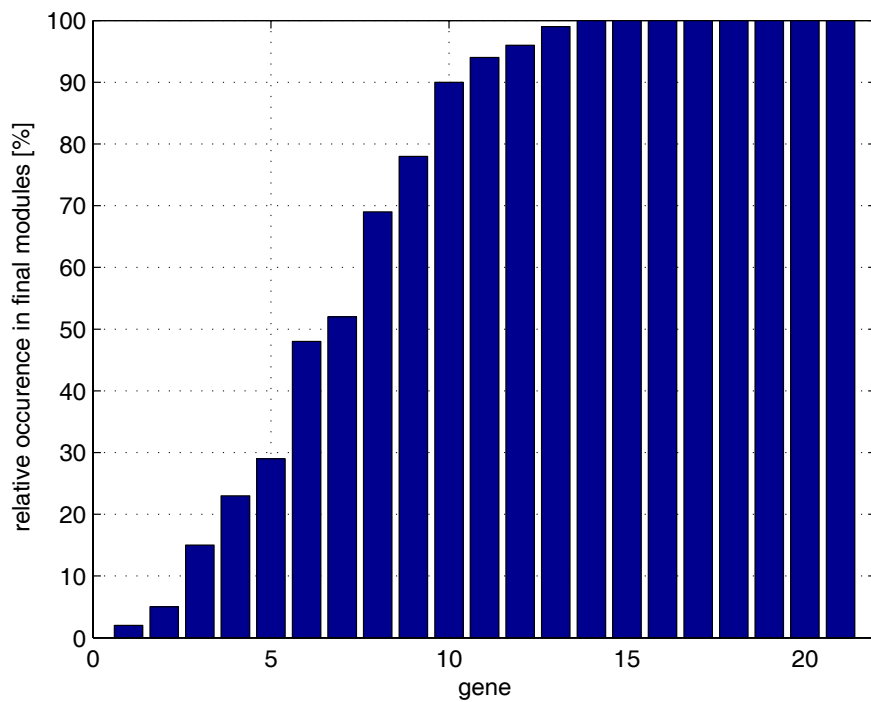


# EC for Data Integration - Bleuler et al. (work in progress)

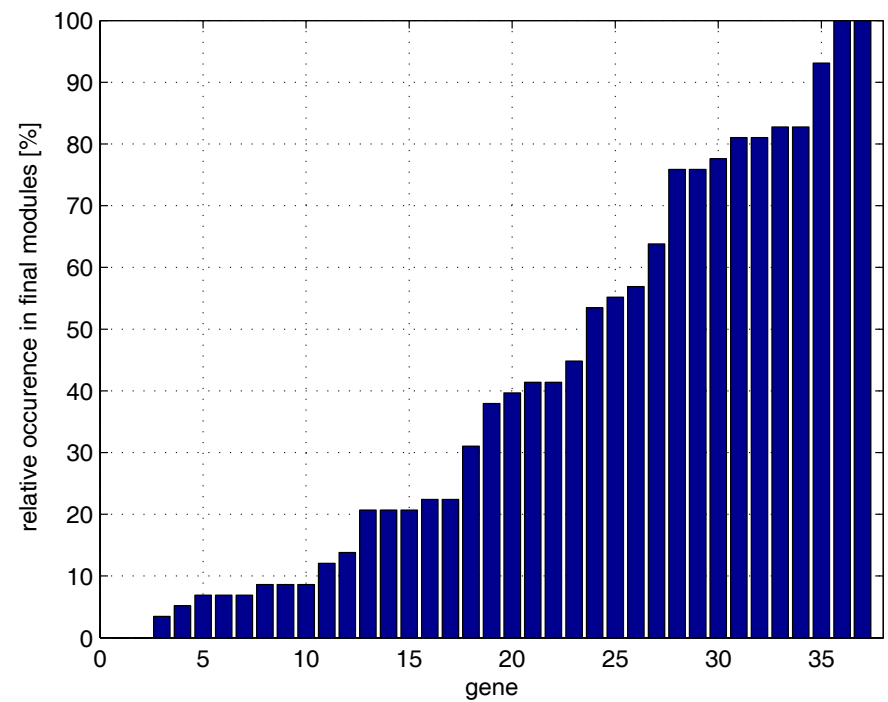


# EC for Data Integration - Bleuler et al. (work in progress)

## Similarity of the Clusters on the Front



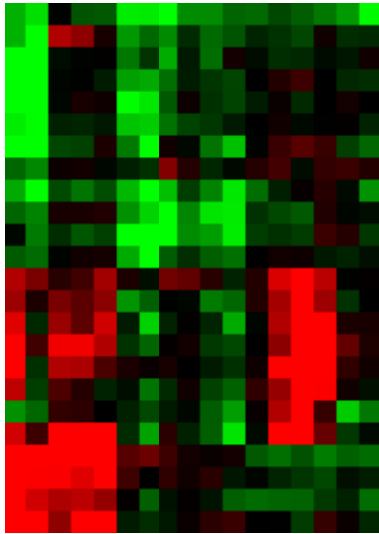
gene expression – gene expression



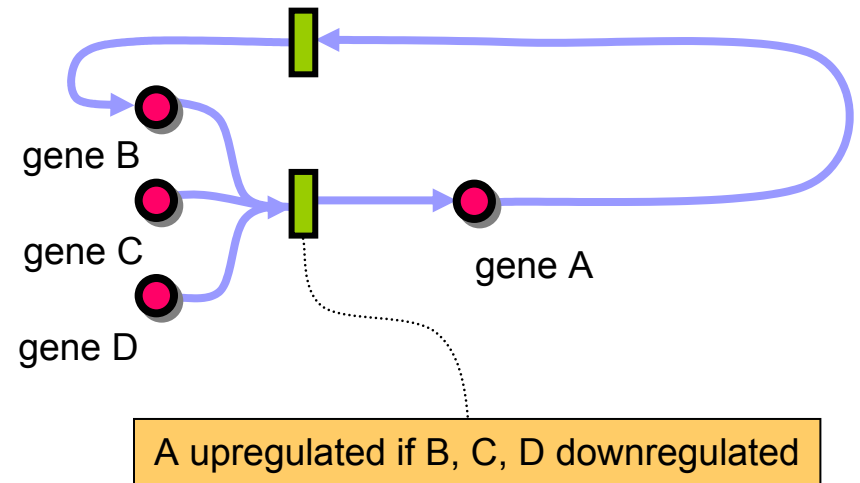
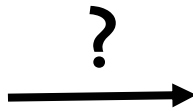
gene expression - PPI

# Network Inference

## Goal



high throughput data



network model

## Approaches

- network analysis (structure, robustness, etc.)
- inference of network topology (Bayesian networks, Gaussian Graphical Models, etc.)
- inference of network function (Boolean networks, differential equations, etc.)

## Challenges

- underdetermined problem
- noisy data
- experiment design

# EC for Network Inference

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## Network Models

- S-systems [3, 6, 7]
- Petri nets [4]
- electronic circuit [1]
- differential equations [3]
- real valued matrix [5]

- [13] J. Koza et al., **Reverse Engineering of Metabolic Pathways from Observed Data Using Genetic Programming**, PSB, 2001
- [14] S. Ando et al., **Modeling Genetic Network by Hybrid GP**, CEC 2002
- [15] S Kikuchi et al., **Dynamic Modeling of Genetic Networks Using Genetic Algorithm and S-System**, Bioinformatics, 2003
- [16] J. Kitagawa and H. Iba., **Identifying Metabolic Pathways and Gene Regulation Networks with Evolutionary Algorithms**, chapter in “Evolutionary Computation in Bioinformatics”, Morgan Kaufmann, 2003
- [17] D. Corne and C. Pridgeon, **Investigating Issues in Reconstructability of Genetic Regulatory Networks**, CEC 2004
- [18] S. Kimura et al., **Inference of S-system models of genetic networks using a cooperative coevolutionary algorithm**, Bioinformatics, 2005
- [19] D.-Y. Cho. et al., **Identification of Biochemical Networks by S-Tree Based Genetic Programming**, Bioinformatics, 2006

# EC for Network Inference – Koza et al. [13]

## Individual

- chemical reaction network
- modeled as electronic circuit
- represented as GP tree

## Objective Function

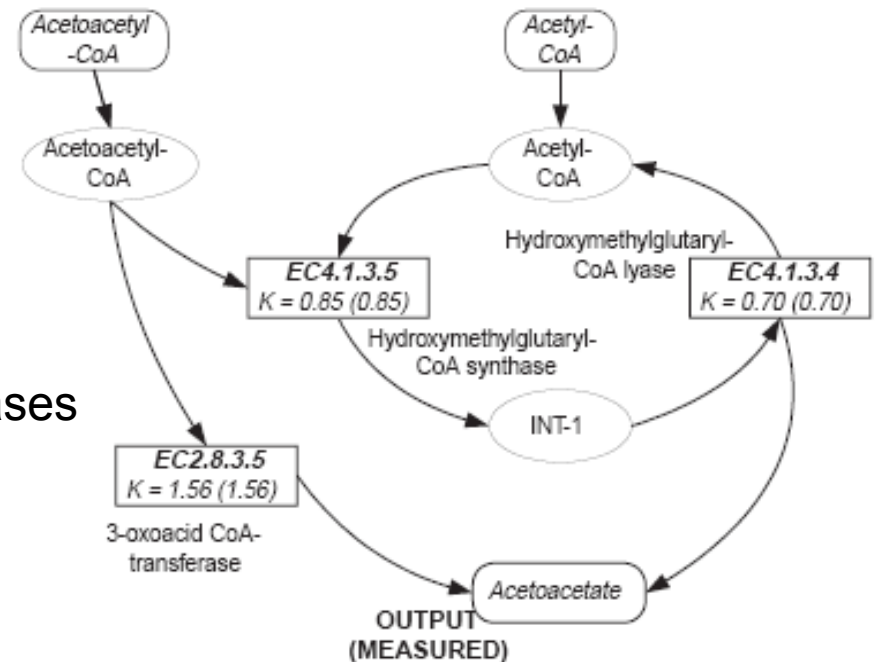
- comparing predicted and measured concentration of end product
- sum of absolute differences for test cases
- evaluated using SPICE

## Optimizer

- GP
- popsize 100'000

## Results

- input: E-cell simulation of phospholipid cycle (4 reactions) and synthesis of ketone bodies (3 reactions)
- good recovery of network topology and reaction rates



# EC for Network Inference – Cho et al. [19]

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## Individual

- biochemical reaction network or gene regulatory network
- modeled as S-tree
- represented as GP tree

## Objective Function

- comparing prediction to measurement on all time points and all substances
- sum of relative squared errors

## Optimizer

- GP
- local hill climbing

## Results

- 1. input: simulation of artificial networks modeled as S-systems
- 1. results: good recovery of network topology and parameters
- 2. input: gene expression from SOS DNA repair in *E. coli* (6 genes)
- 2. results: all but one known interaction recovered (in 35 h).

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### 3. Status Quo and Future Trends



# Status Quo

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## **Advantages of EC Approach**

- flexible
- global search method
- multiobjective

## **Open Problems**

- benchmark problems missing
- little comparison with non-EA methods
- no common methodology

# Future Trends

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## Biology and Measurements

- more data (more genomes, transcriptomes and proteomes)
- more data types (tiling arrays, synthetic lethal, etc.)
- more specific measurements (towards single cell analytics)
- more formalized information about experiments

## Computational

### Data Integration of ...

- different qualities (accuracy)
- different data types (proteomics, metabolomics, etc.)
- different scales
- different precision (qualitative vs. quantitative)